

Structure, Function and Regulation of Versican: The Most Abundant Type of Proteoglycan in the Extracellular Matrix

Fattah Sotoodehnejadnematlahi¹ and Bernard Burke²

¹Department of Regenerative Biomedicine at Cell Science Research Center, Royan Institute for Stem Cell Biology and Technology, ACECR, Tehran, Iran

²Department of Infection, Immunity and Inflammation, University of Leicester, Leicester, LE1 9HN, UK

Received: 20 May 2012; Accepted: 26 Feb. 2013

Abstract- One of the main members of the large aggregating proteoglycans (PGs) family is versican which is able to bind to hyaluronate. Versican is a chondroitin sulfate proteoglycan and is a key ingredient of the extracellular matrix. Due to its widespread expression in the body, versican is involved in cell adhesion, proliferation and migration. Induced expression of versican is often observed in tissues such as breast, brain, ovary, gastrointestinal tract, prostate, and melanoma. In addition, versican has important role in development. For example, versican conducts the embryonic cell migration which is essential in the formation of the heart and outlining the path for neural crest cell migration. Several studies in the past decade up to now have shown that versican produced by mononuclear cells has an important role in wound healing and blood vessel formation and suggested that it promotes tumorigenesis and angiogenesis. In this mini-review, we summarise and discuss the role of versican in healthy and pathological tissues and suggest the possible function of transcription factors and signalling pathway in regulation of versican.

© 2013 Tehran University of Medical Sciences. All rights reserved.

Acta Medica Iranica, 2013; 51(11): 740-750.

Keywords: Extracellular matrix; Proteoglycans; Versican

Introduction

The extracellular matrix (ECM) which provides structural support for organs and tissues (1) is also forming the basement membranes for the cell layers (2) and cell migration (3). ECM is composed of collagens and elastic fibres which are embedded in a viscoelastic gel that comprises proteoglycans (*e.g.* versican and hyaluronan), glycoproteins and water (4,5). ECM forms a complex, three-dimensional network among the cells of different tissues in an organ-specific manner (6) and plays vital roles in the differentiation, proliferation and survival of cells (7-9).

Proteoglycans are the main components of ECM, and are characterised by a protein portion (core protein) and one or more unbranched, long and negatively charged polysaccharide chains called glycosaminoglycans (GAG) which are covalently attached to the core protein (10,11). Depending upon the nature of the GAG chains, proteoglycans can be categorised as heparan sulphate proteoglycans, chondroitin sulphate proteoglycans

(CSPGs) and dermatan sulphate proteoglycans, or keratan sulphate proteoglycans (12,13). Of these, the CSPGs such as versican are the most abundant type of proteoglycan in the ECM of mammalian tissues (14,15).

Versican structure

Versican is a large chondroitin sulphate proteoglycan which is a major component of the ECM (16). Versican is transcribed from a single gene which is localized on chromosome 5q 12-14 in the human genome and extends over 90 kb (17) which is divided into 15 exons which range in size from 76 to 5262 bp (16). The alternative mRNA-splicing of these exons gives rise to four different versican isoforms which are distinguished by different core-middle regions (18). Versican is comprised of three domains. The amino terminal G1 domain interacts with a GAG called hyaluronan present in the extracellular matrix (14). The carboxyl terminal domain of versican is called the G3 domain and it contains a C-type lectin binding domain, two epidermal growth factor repeats and a complement regulatory

Corresponding Author: Fattah Sotoodehnejadnematlahi

Department of Stem Cells and Developmental Biology, Royan Institute for Stem Cell Biology and Technology, Banihashem Sq., Banihashem St., Resalat highway, P.O. Box: 19395-4644, Tehran, Iran

Tel: +98 21 22306485, Fax: +98 21 22310406, E-mail: sotoodeh@Royaninstitute.org, fattah212@gmail.com

region. The versican core protein contains the GAG attachment region and the chondroitin sulphate chains extend from this region of the protein (Figure 1) (14,19,20).

Versican isoforms

The central area of versican is encoded by two exons that specify chondroitin sulphate attachment regions (14). RNA splicing of these two exons generates four isoforms of versican named V0, V1, V2 and V3 core

protein molecular weight of about 370 kDa, 263 kDa, 180 kDa, and 74 kDa, respectively (18). V0, the largest versican isoform, is encoded by exons 7 and 8 and contains the GAG- α and β domains. The V1 isoform contains GAG- β attachment domain which is encoded by exon 8 (lacking exon 7) whereas the V2 isoform contains a GAG- α domain which is encoded by exon 7 (lacking exon 8) (22). V3 does not include either exon 7 or 8 and consequently has no GAG attachment sites (Figure 2) (19,22).

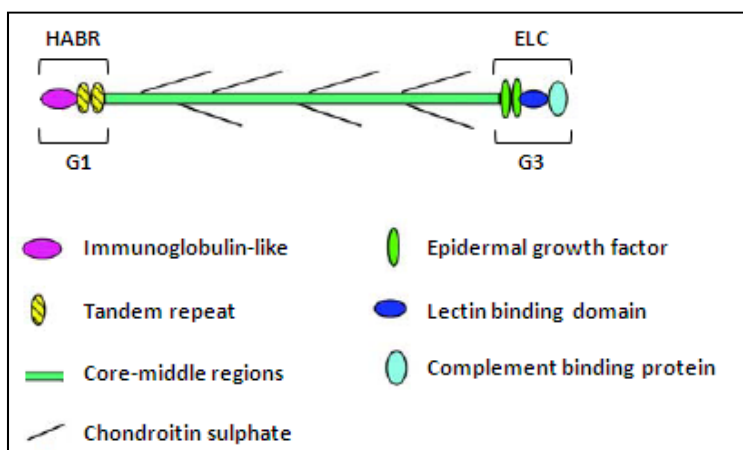


Figure 1. Schematic model of versican structure. Versican contains globular domains at the amino terminus (G1) and carboxyl terminus (G3). The G1 domain is composed of an immunoglobulin-like motif, followed by two proteoglycan tandem repeats which bind hyaluronan (HABR; hyaluronan binding region). The G3 domain contains two epidermal growth factor-like repeats, a carbohydrate recognition domain (a lectin-binding domain) and complement binding protein (ELC). The core-middle region of versican contains GAG attachment regions that are encoded by exons 7 and 8 which give rise to four different versican isoforms. GAG chondroitin sulphate chains extend from the core protein (21).

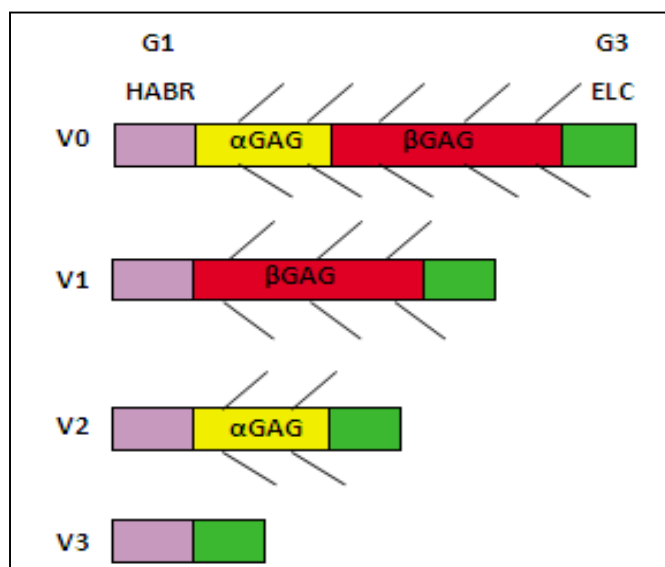


Figure 2. Cartoon of versican isoforms generated by alternative splicing of the mRNA transcript. Different colours show specific domains in the gene. G1 and G3 are shown at the amino and carboxyl termini respectively. Purple = hyaluronan binding region (HABR); yellow = α GAG exon; red = β GAG exon and green = epidermal growth factor repeats (E), a lectin binding domain (L) and a complement regulatory region (C). The glycosaminoglycan chondroitin sulphate chains are shown as (/)

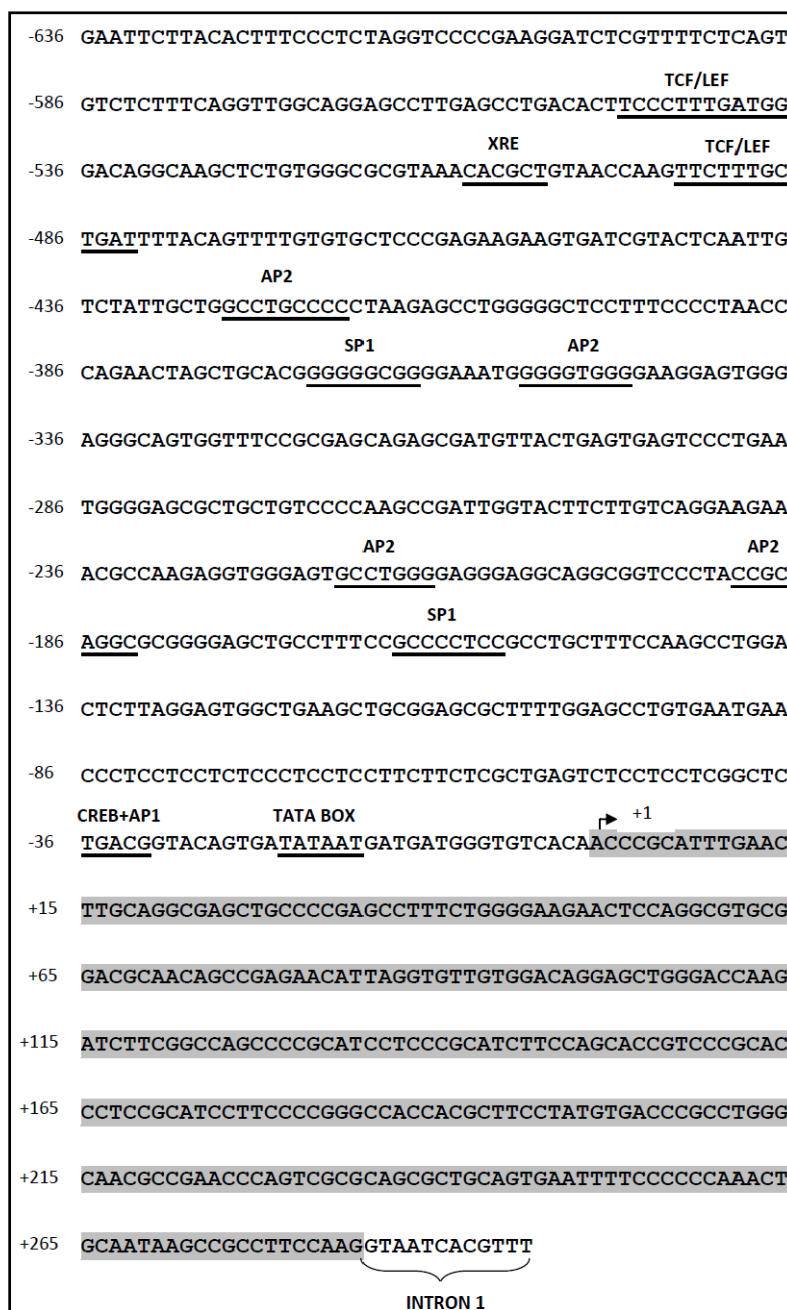


Figure 3. Sequence analysis of versican gene promoter and exon 1. Naso *et al.*, revealed a typical TATA box located approximately 16 bp upstream of the transcription start site and binding sites for a number of transcription regulatory factors including CREB at position -36 bp, SP-1 at position -370 bp, XRE at position -509 bp and AP-2 at positions -426 bp, -355 bp, -218 bp and -190bp respectively (16). Rahmani *et al.*, showed two binding sites for TCF/LEF at positions -546 bp and -492 bp respectively (29). Also further investigation by Domenzain *et al.*, revealed one binding site for SP-1 at position -166 bp and one binding site for AP-1 at position -36 bp. Positions of all transcription factors are relative to the transcription start site (30). The sequence of exon 1 is shaded. CREB; cAMP response element-binding, AP-1& 2; (activator protein 1&2), XRE; xenobiotic responsive element, TCF/LEF; T-cell factor/lymphoid enhancer factor.

An interesting study on human adult tissues showed that all four versican isoforms are transcribed in more than 50% of tissues, although, intriguingly, only the V1

isoform is expressed in liver and spleen (23). Another study suggested that V1 versican enhances cell proliferation and protects mouse fibroblast cell lines

from apoptosis whereas the V2 isoform exhibits opposite activities by inhibiting cell proliferation (24).

In addition, it has been reported that V0 and V1 are the predominantly expressed isoforms in tumours suggesting that these isoforms are mainly involved in tumour development (25,26). In contrast, over-expression of V3 (the smallest versican isoform which consists of only the G1 and G3 domains) markedly reduced cell growth in melanoma cancer cells, suggesting a role for V3 versican as an inhibitor of tumour growth (27,28).

Versican promoter

The transcription start site for the versican gene was first identified by Naso *et al.*, in 1994 (16). This study reported that the versican promoter contains a typical TATA box located approximately 16 base pairs upstream of the transcription start site (Figure 3). Transient transfection of a reporter construct driven by an 876-bp (-632/+240) fragment of the versican promoter showed significant expression in HeLa cells and IMR-90 embryonic lung fibroblasts. Furthermore, deletion constructs of the 876-bp confirmed that the human versican 5'-flanking sequence contains promoter, enhancer and repressor elements which are able to drive the expression of the versican reporter gene in different cells (14). In addition, sequence analysis has revealed potential binding sites for several transcription factors in the 876 bp versican promoter region including CREB (14), T-cell factor/ lymphoid enhancer-binding factor (TCF/LEF) (29), AP-1 and SP1 (30) (Figure 3).

Versican function

Versican is a main component of the ECM where its hygroscopic properties create a loose and hydrated matrix which is necessary to support key events in development (31,32). Versican is found in a variety of tissues including the brain (33) and skin (34). Increased expression of versican is also observed in sites of tissue injury (35) and in cancers including breast (36), ovarian (37), gastrointestinal tract (38), prostate (39), brain (40), cervical (41) and melanoma (42). Several reports have also highlighted the role of versican in wound healing (23), angiogenesis, tumour growth (43) and in vascular diseases, especially atherosclerosis (44,45). It has been demonstrated that versican binds low-density lipoprotein (LDL) particles and it is believed that accumulation of versican in blood vessels promotes extracellular lipoprotein retention, suggesting roles in lipid accumulation, inflammation and thrombosis (46,47). Due to versican's structural composition and its

widespread expression in the body it is able to regulate cell adhesion and survival, cell proliferation, cell migration and ECM assembly (48) that the key studies in these areas are reviewed below.

Cell adhesion

Early studies reported that most chondroitin sulphate proteoglycans may be considered as anti-adhesion molecules for the regulation of cell adhesion to the substratum, which is essential for various cell and tissue functions (49,50). Different studies presented evidence suggesting that this inhibitory activity could be due to the G1 domain of versican (49,51). They showed that selective exclusion of versican from podosomes of cultured human osteosarcoma cells suppresses the malignant cell-adhesive phenotype, suggesting that versican can act as an anti-adhesive molecule (52). However there is evidence that the carboxy-terminal domain of versican interacts with the β 1 integrin of brain tumour cells leading to the activation of focal adhesion kinase (FAK), promoting cell adhesion and protecting the cell from apoptosis (53,54).

Interaction of versican with selectins and chemokines has been studied. It has been shown that versican binds to selectins, adhesion molecules on the surface of activated endothelial cells, through its chondroitin sulphate chains (55). In addition, versican has been shown to bind secondary lymphoid tissue chemokine (SLC) through chondroitin sulphate chains and this binding tends to down-regulate chemokine function for recruitment of lymphocytes (56). Taken together the data suggest that versican, which is induced in inflammatory conditions such as arthritis (57), asthma and lung disease (58,59), may regulate inflammation by regulating interaction with selectins and chemokines (56).

Cell proliferation

Abundant expression of versican in fast growing tissues and cells suggested a key role for versican in cell proliferation (60,61). For example high expression of versican is detected in the loose connective tissue of various organs including the central and peripheral nervous system (60), blood vessels (62), dermis and in the proliferative zone of the epidermis (34).

Versican is also involved in the proliferation of smooth muscle cells (SMC) (63,64). Several studies have reported proteins such as platelet-derived growth factor (PDGF) and transforming growth factor- β 1 (TFG- β 1) increase versican synthesis in arterial smooth muscle cells (ASMCs) (63,65). It

Versican; structure, function and regulation

was demonstrated that increases in versican and the associated protein hyaluronan in response to PDGF and TGF- β 1 cause increases in the pericellular matrix of the cells and expansion of the ECM that is required for the proliferation and migration of these cells (66). In addition it was shown that proliferation of ASMCs treated with PDGF is blocked by inhibition of the formation of versican-hyaluronan complex which serves as an important mechanism for controlling cell shape and cell division (67).

Other studies have suggested a role for versican in cell proliferation through its two epidermal growth factor sequences in the G3 domain of the molecule (68, 69). These studies showed that expression of the G3 domain of versican promotes proliferation in NIH3T3 fibroblasts cells whereas this effect can be inhibited by removing EGF motifs in the versican G3 domain (68). These results suggested that the EGF-like motifs in the versican G3 domain may promote cell proliferation through direct or indirect interaction with the EGF receptor (EGFR) (70).

Cell migration

Controversial studies have demonstrated that versican is widely expressed at both mRNA and protein level in neural crest pathways and influences neural cell migration (71) whereas a number of other studies have shown that versican prevents migration of these cells (35,72). These contradictory findings are believed to be due to different versican isoforms which differ in the core-middle region (73). Some studies have investigated the role of versican in the nervous system and axonal outgrowth (74-76). These studies showed that chondroitin sulphate (CS) chains of versican isotype V2 are involved in inhibiting axonal outgrowth and migration of the mature nervous system. The role of versican in axonal migration was investigated by Asher *et al.*, in 2002 (77) who showed up-regulation of versican following central nervous system (CNS) injury and suggested these changes in versican regulation are associated with the failure of nerves to regenerate.

As mentioned earlier, the G3 domain of versican can interact with integrin β 1 which is able to form clusters with EGF receptors (78). Growing evidence indicates that interaction of integrins with EGF receptors induces down-stream signal to extracellular regulated kinase (ERK) which is crucial in regulating a range of cell activities, such as migration (79-81). Also, the role of versican in the migration of embryonic cells in the

development of the heart has been studied (82, 83). It has been reported that versican mRNA and protein is strongly expressed during the development of mouse heart suggesting a key role for versican in cardiac development (82).

Furthermore other studies have reported that the G3 domain of versican directly interacts with fibronectin, another extracellular matrix glycoprotein (84) and showed that formation of a complex of versican G3 domain and fibronectin with VEGF can enhance endothelial cell migration which this process was reversed by removal of the complex (43). This study and other investigation by Wijelath *et al.*, in 2002 (85) indicated that expression of versican G3 enhanced brain tumour growth, suggesting the role of versican G3 fragment on promoting angiogenesis and tumour growth that it suggests targeting versican G3 fragments may help to develop a new approach for anticancer and anti-angiogenic therapies.

Extracellular matrix assembly

Versican interacts with different ECM molecules and has been reported to have an important role during ECM assembly (21). Possibly the best known is a specific interaction between the G1 domain of versican and hyaluronan (86). Hyaluronan is a large polysaccharide in the ECM and is able to create a lattice structure which may regulate cell adhesion and migration (87). Versican binds hyaluronan and this binding requires the double tandem repeat present in the G1 domain of versican (88,89).

Versican regulation

Signal transduction pathways

The signalling pathways which modulate versican synthesis are not fully understood although studies on the intracellular pathways have reported that PDGF-stimulated versican expression in arterial smooth muscle cells (SMC) is regulated by endogenous tyrosine kinase activity of the PDGF receptor which up-regulates versican synthesis at both mRNA and protein levels in vascular smooth muscle cells (63,90). In addition, another study has suggested the role of protein kinase C (PKC) and ERK in the PDGF stimulated versican gene expression in non-human ASMC (91).

An interesting study by Rahmani *et al.*, in 2005 (29) demonstrated the role of the PI3-K/ PKB (Protein kinase B) pathway in versican expression in SMC. They suggested that phosphorylation and inactivation of glycogen synthase kinase-3 β (GSK-3 β), a downstream

effector of PI3K/PKB (92), leads to activation and nuclear accumulation of β -catenin which binds to the TCF/LEF transcription factors in the versican promoter and then increases versican transcription in SMC (Figure 4) (29). Further investigation by this group using a specific inhibitor of the PI3-Kinase pathway, LY294002, showed inhibited activation of downstream PKB and resulted in significant inhibition of versican promoter reporter activity and versican mRNA expression in SMC. A similar mechanism has been observed for other genes such as MMP-7 (93) and VEGF (94), which have also been shown to be targeted by PI3-K/PKB and up-regulated by formation of a β -catenin TCF/LEF complex.

Transcription factors

Analysis of the versican promoter by Rahmani *et al.*, in 2005 (29) revealed two putative binding sites for TCF/LEF transcription factors at positions -546 and -492 bp relative to the transcription start site (Figure 3). They showed that site-directed mutagenesis of the TCF

sites in the versican promoter markedly diminished reporter luciferase activity in SMC. Furthermore, electrophoretic mobility shift assay (EMSA) and supershift assays revealed that the β -catenin/TCF transcription factor complex is essential for versican expression in SMC (29).

A recent study by Domenzain *et al.*, identified several other transcriptional regulatory elements including AP1, SP1 and AP2 on a 620 bp (-618/+2 relative to the transcriptional start site) in a proximal versican promoter reporter construct (30). This study demonstrated that mutagenesis of the AP-1 site at position -36 bp completely abolished versican promoter activity in human melanoma cell lines. Also further investigation by EMSA confirmed the importance of the AP-1 binding site for versican promoter transcription in these cell lines. In addition, versican promoter activity in a TCF/LEF mutated construct was reduced by half, suggesting that versican expression is also up-regulated via the β -catenin/TCF pathway in human melanoma cell lines (30).

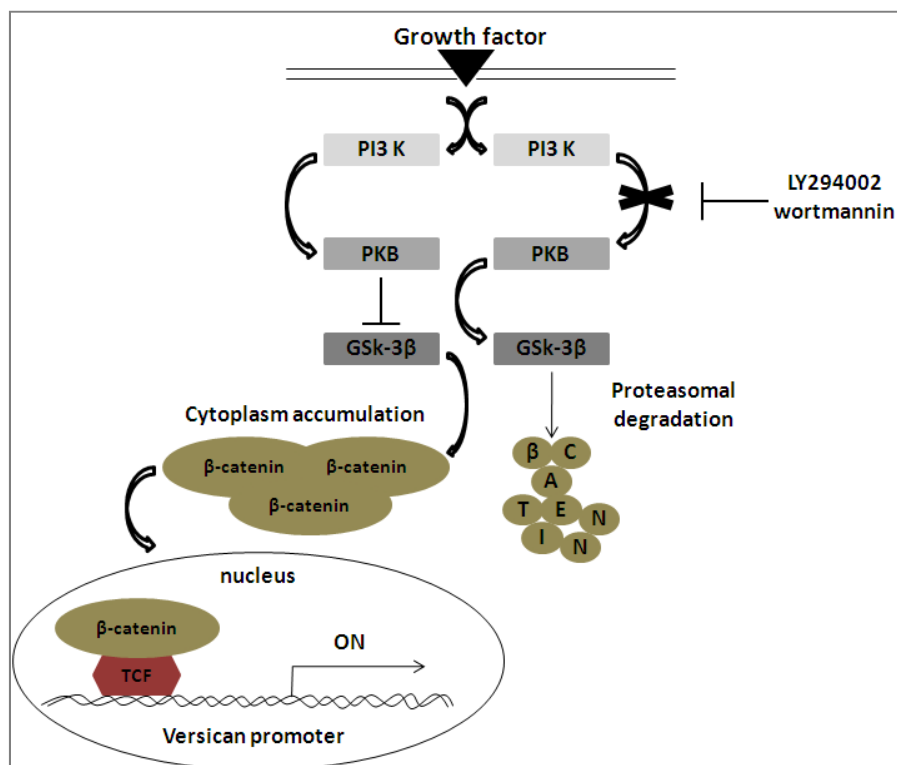


Figure 4. Schematic model of versican promoter regulation via PI3/PKB signalling and β -catenin TCF transcription factor complex suggested by Rahmani *et al.* (29). Activation of PI3K signalling by growth factors leads to phosphorylation and inactivation of GSK-3 β which results in β -catenin cytoplasmic accumulation and subsequent translocation to the nucleus. Nuclear accumulation of β -catenin leads to complex formation with TCF/LEF transcription factors and transactivation of TCF/LEF target genes. Specific inhibitors of the PI3K signalling pathway such as LY294002 and wortmannin lead to activation of GSK-3 β and subsequently to β -catenin degradation.

Versican; structure, function and regulation

Yoon *et al.*, investigated the role of the transcription factor p53 in versican gene expression in a broad range of human carcinoma cell lines (95). P53 is a transcription factor involved in important cellular processes such as cell cycle checkpoint regulation, DNA damage and apoptosis (96,97). Oligonucleotide-array gene expression analysis of human carcinoma cell lines by Yoon *et al.*, demonstrated high expression of the versican gene in wild type p53 (p53 +/+) cells but lower expression in p53 -/- cells, suggesting that versican is a direct target of p53. Further investigation using wild type p53 over-expression in p53-null cells transfected with versican promoter reporter constructs showed 200-fold increases in luciferase activity in comparison with the control plasmid. EMSA and super-shift assays confirmed the interaction of p53 protein and the versican p53 binding site (95). In conclusion, in the present review, I provided evidence about versican structure, regulation and its role in normal tissue and tumour growth. In addition, this review discussed the function of versican as a key factor in inflammation through interactions with adhesion molecules on the surfaces of inflammatory leukocytes and interactions with chemokines that are involved in recruiting inflammatory cells. A better understanding of how versican is regulated will hopefully be helpful for the development of future therapies for a range of different disease such as vascular disorders, including atherosclerosis, where versican accumulation plays a key role.

References

1. Badylak SF. The extracellular matrix as a scaffold for tissue reconstruction. *Semin Cell Dev Biol* 2002;13(5):377-83.
2. Daley WP, Peters SB, Larsen M. Extracellular matrix dynamics in development and regenerative medicine. *J Cell Sci* 2008;121(Pt3):255-64.
3. Werb Z. ECM and cell surface proteolysis: regulating cellular ecology. *Cell* 1997;91(4):439-42.
4. Berrier AL, Yamada KM. Cell-matrix adhesion. *J Cell Physiol* 2007;213(3):565-73.
5. Campbell NE, Kellenberger L, Greenaway J, Moorehead RA, Linnerth-Petrik NM, Petrik J. Extracellular matrix proteins and tumor angiogenesis. *J Oncol* 2010; 586905.
6. Adams JC, Watt FM. Regulation of development and differentiation by the extracellular matrix. *Development* 1993;117(4):1183-98.
7. Boudreau N, Bissell MJ. Extracellular matrix signaling: integration of form and function in normal and malignant cells. *Curr Opin Cell Biol* 1998;10(5):640-6.
8. Streuli C. Extracellular matrix remodelling and cellular differentiation. *Curr Opin Cell Biol* 1999;11(5):634-40.
9. Van Horsen J, Dijkstra CD, de Vries HE. The extracellular matrix in multiple sclerosis pathology. *J Neurochem* 2007;103(4):1293-301.
10. Hardingham TE, Fosang AJ. Proteoglycans: many forms and many functions. *FASEB J* 1992;6(3):861-70.
11. Iozzo RV, Murdoch AD. Proteoglycans of the extracellular environment: clues from the gene and protein side offer novel perspectives in molecular diversity and function. *FASEB J* 1996;10(5):598-614.
12. Iozzo RV. Matrix proteoglycans: from molecular design to cellular function. *Annu Rev Biochem* 1998;67:609-52.
13. Kresse H, Schonherr E. Proteoglycans of the extracellular matrix and growth control. *J Cell Physiol* 2001;189(3):266-74.
14. Zimmermann DR, Ruoslahti E. Multiple domains of the large fibroblast proteoglycan, versican. *EMBO J* 1989;8(10):2975-81.
15. Carulli D, Laabs T, Geller HM, Fawcett JW. Chondroitin sulfate proteoglycans in neural development and regeneration. *Curr Opin Neurobiol* 2005;15(2):116-20.
16. Naso MF, Zimmermann DR, Iozzo RV. Characterization of the complete genomic structure of the human versican gene and functional analysis of its promoter. *J Biol Chem* 1994;269(52):32999-3008.
17. Iozzo RV, Naso MF, Cannizzaro LA, Wasmuth JJ, McPherson JD. Mapping of the versican proteoglycan gene (CSPG2) to the long arm of human chromosome 5 (5q12-5q14). *Genomics* 1992;14(4):845-51.
18. Dours-Zimmermann MT, Zimmermann DR. A novel glycosaminoglycan attachment domain identified in two alternative splice variants of human versican. *J Biol Chem* 1994;269(52):32992-8.
19. Zako M, Shinomura T, Ujita M, Ito K, Kimata K. Expression of PG-M(V3), an alternatively spliced form of PG-M without a chondroitin sulfate attachment in region in mouse and human tissues. *J Biol Chem* 1995;270(8):3914-8.
20. Yang BL, Cao L, Kiani C, Lee V, Zhang Y, Adams ME, Yang BB. Tandem repeats are involved in G1 domain inhibition of versican expression and secretion and the G3 domain enhances glycosaminoglycan modification and product secretion via the complement-binding protein-like motif. *J Biol Chem* 2000;275(28):21255-61.
21. Wu YJ, La Pierre DP, Wu J, Yee AJ, Yang BB. The interaction of versican with its binding partners. *Cell Res* 2005;15(7):483-94.

22. Lemire JM, Braun KR, Maurel P, Kaplan ED, Schwartz SM, Wight TN. Versican/PG-M isoforms in vascular smooth muscle cells. *Arterioscler Thromb Vasc Biol* 1999;19(7):1630-9
23. Cattaruzza S, Schiappacassi M, Ljungberg-Rose A, Spessotto P, Perissinotto D, Morgelin M, Mucignat MT, Colombatti A, Perris R. Distribution of PG-M/versican variants in human tissues and de novo expression of isoform V3 upon endothelial cell activation, migration, and neoangiogenesis in vitro. *J Biol Chem* 2002;277(49):47626-35.
24. Sheng W, Wang G, Wang Y, Liang J, Wen J, Zheng PS, Wu Y, Lee V, Slingerland J, Dumont D, Yang BB. The roles of versican V1 and V2 isoforms in cell proliferation and apoptosis. *Mol Biol Cell* 2005;16(3):1330-40.
25. Touab M, Villena J, Barranco C, Arumi-Uria M, Bassols A. Versican is differentially expressed in human melanoma and may play a role in tumor development. *Am J Pathol* 2002;160(2):549-57.
26. Arslan F, Bosserhoff AK, Nickl-Jockschat T, Doerfelt A, Bogdahn U, Hau P. The role of versican isoforms V0/V1 in glioma migration mediated by transforming growth factor-beta2. *Br J Cancer* 2007;96(10):1560-8.
27. Serra M, Miquel L, Domenzain C, Docampo MJ, Fabra A, Wight TN, Bassols A. V3 versican isoform expression alters the phenotype of melanoma cells and their tumorigenic potential. *Int J Cancer* 2005;114(6):879-86.
28. Hernandez D, Miquel-Serra L, Docampo M.J., Marco-Ramell, A., Cabrera, J., Fabra, A., and Bassols, A. V3 versican isoform alters the behavior of human melanoma cells by interfering with CD44/ErbB-dependent signaling. *J Biol Chem* 2010; 286(2): 1475-1485.
29. Rahmani M, Read JT, Carthy JM, McDonald PC, Wong BW, Esfandiarei M, Si X, Luo Z, Luo H, Rennie PS, McManus BM. Regulation of the versican promoter by the beta-catenin-T-cell factor complex in vascular smooth muscle cells. *J Biol Chem* 2005;280(13):13019-28.
30. Domenzain C, Hernandez D, Miquel-Serra L, Docampo MJ, Badenas C, Fabra A, Bassols A. Structure and regulation of the versican promoter: the versican promoter is regulated by AP-1 and TCF transcription factors in invasive human melanoma cells. *J Biol Chem* 2009;284(18):12306-17.
31. Lemire JM, Merrilees MJ, Braun KR, Wight TN. Overexpression of the V3 variant of versican alters arterial smooth muscle cell adhesion, migration, and proliferation in vitro. *J Cell Physiol* 2002;190(1):38-45.
32. Hinek A, Braun KR, Liu K, Wang Y, Wight TN. Retrovirally mediated overexpression of versican v3 reverses impaired elastogenesis and heightened proliferation exhibited by fibroblasts from Costello syndrome and Hurler disease patients. *Am J Pathol* 2004;164(1):119-31.
33. Perides G, Rahemtulla F, Lane WS, Asher RA, Bignami A. Isolation of a large aggregating proteoglycan from human brain. *J Biol Chem* 1992;267(33):23883-7.
34. Zimmermann DR, Dours-Zimmermann MT, Schubert M, Bruckner-Tuderman L. Versican is expressed in the proliferating zone in the epidermis and in association with the elastic network of the dermis. *J Cell Biol* 1994(5);124:817-25.
35. Landolt RM, Vaughan L, Winterhalter KH, Zimmermann, DR. Versican is selectively expressed in embryonic tissues that act as barriers to neural crest cell migration and axon outgrowth. *Development* 1995;121(8):2303-12.
36. Kischel P, Waltregny D, Dumont B, Turtoi A, Greffe Y, Kirsch S, De Pauw E, Castronovo V. Versican overexpression in human breast cancer lesions: known and new isoforms for stromal tumor targeting. *Int J Cancer* 2010;126(3):640-50.
37. Voutilainen K, Anttila M, Sillanpaa S, Tammi R, Tammi M, Saarikoski S, Kosma VM. Versican in epithelial ovarian cancer: relation to hyaluronan, clinicopathologic factors and prognosis. *Int J Cancer* 2003;107(3):359-64.
38. Theocharis AD. Human colon adenocarcinoma is associated with specific post-translational modifications of versican and decorin. *Biochim Biophys Acta* 2002; 1588(2): 165-172.
39. Sakko AJ, Ricciardelli C, Mayne K, Suwiwat S, LeBaron RG, Marshall VR, Tilley WD, Horsfall DJ. Modulation of prostate cancer cell attachment to matrix by versican. *Cancer Res* 2003;63(16):4786-91.
40. Schwartz NB, Domowicz M. Proteoglycans in brain development. *Glycoconj J* 2004;21(6):329-41.
41. Kodama J, Hasengaowa Kusumoto T, Seki N, Matsuo T, Nakamura K, Hongo A, Hiramatsu Y. Versican expression in human cervical cancer. *Eur J Cancer* 2007;43(9):1460-6.
42. Domenzain C, Docampo MJ, Serra M, Miquel L, Bassols A. Differential expression of versican isoforms is a component of the human melanoma cell differentiation process. *Biochim Biophys Acta* 2003;1642(1-2): 107-14.
43. Zheng PS, Wen J, Ang LC, Sheng W, Vilorio-Petit A, Wang Y, Wu Y, Kerbel RS, Yang BB. Versican/PG-M G3 domain promotes tumor growth and angiogenesis. *FASEB J* 2004;18(6):754-6.
44. Talusan P, Bedri S, Yang S, Kattapuram T, Silva N, Roughley PJ, Stone JR. Analysis of intimal proteoglycans in atherosclerosis-prone and atherosclerosis-resistant human arteries by mass spectrometry. *Mol Cell Proteomics* 2005;4(9):1350-7.

Versican; structure, function and regulation

45. Kenagy RD, Plaas AH, Wight TN. Versican degradation and vascular disease. *Trends Cardiovasc Med* 2006; 16(6): 209-215.
46. Olin KL, Potter-Perigo S, Barrett PH, Wight TN, Chait A. Lipoprotein lipase enhances the binding of native and oxidized low density lipoproteins to versican and biglycan synthesized by cultured arterial smooth muscle cells. *J Biol Chem* 1999;274(49):34629-36.
47. Wight TN, Merrilees MJ. Proteoglycans in atherosclerosis and restenosis: key roles for versican. *Circ Res* 2004;94(9):1158-67.
48. Ricciardelli C, Sakko AJ, Ween MP, Russell DL, Horsfall DJ. The biological role and regulation of versican levels in cancer. *Cancer Metastasis Rev* 2009; 28(1-2): 233-245.
49. Yamagata M, Suzuki S, Akiyama SK, Yamada KM, Kimata K. Regulation of cell-substrate adhesion by proteoglycans immobilized on extracellular substrates. *J Biol Chem* 1989;264(14):8012-8.
50. Yamagata M, Saga S, Kato M, Bernfield M, Kimata K. Selective distributions of proteoglycans and their ligands in pericellular matrix of cultured fibroblasts. Implications for their roles in cell-substratum adhesion. *J Cell Sci* 1993;106(Pt 1):55-65.
51. Sugiura, N, Sakurai K, Hori Y, Karasawa K, Suzuki S, Kimata K. Preparation of lipid-derivatized glycosaminoglycans to probe a regulatory function of the carbohydrate moieties of proteoglycans in cell-matrix interaction. *J Biol Chem* 1993;268(21):15779-87.
52. Yamagata M, Kimata K. Repression of a malignant cell-substratum adhesion phenotype by inhibiting the production of the anti-adhesive proteoglycan, PG-M/versican. *J Cell Sci* 1994; 107(Pt 9): 2581-2590.
53. Wu Y, Chen L, Zheng PS, Yang BB. beta 1-Integrin-mediated glioma cell adhesion and free radical-induced apoptosis are regulated by binding to a C-terminal domain of PG-M/versican. *J Biol Chem* 2002;277(14):12294-301.
54. Wu Y, Chen L, Cao L, Sheng W, Yang BB. Overexpression of the C-terminal PG-M/versican domain impairs growth of tumor cells by intervening in the interaction between epidermal growth factor receptor and beta1-integrin. *J Cell Sci* 2004;117(Pt 11):2227-37.
55. Kawashima H, Atarashi K, Hirose M, Hirose J, Yamada S, Sugahara K, Miyasaka M. Oversulfated chondroitin/dermatan sulfates containing GlcA β 1/IdoA α 1-3GalNAc (4,6-O-disulfate) interact with L- and P-selectin and chemokines. *J Biol Chem* 2002;277(15):12921-30.
56. Hirose J, Kawashima H, Yoshie O, Tashiro K, Miyasaka M. Versican interacts with chemokines and modulates cellular responses. *J Biol Chem* 2001;276(7):5228-34.
57. Wight TN, Merrilees MJ. Proteoglycans in atherosclerosis and restenosis: key roles for versican. *Circ Res* 2004;94(9):1158-67.
58. Johnson PR. Role of human airway smooth muscle in altered extracellular matrix production in asthma. *Clin Exp Pharmacol Physiol* 2001;28(3):233-6.
59. Merrilees MJ, Hankin EJ, Black JL, Beaumont B. Matrix proteoglycans and remodelling of interstitial lung tissue in lymphangioliomyomatosis. *J Pathol* 2004;203(2):653-60.
60. Bode-Lesniewska B, Dours-Zimmermann MT, Odermatt BF, Briner J, Heitz PU, Zimmermann DR. Distribution of the large aggregating proteoglycan versican in adult human tissues. *J Histochem Cytochem* 1996;44(4):303-12.
61. Hernandez D, Miquel-Serra L, Docampo MJ, Marco-Ramell A, Cabrera J, Fabra A, Bassols A. V3 versican isoform alters the behavior of human melanoma cells by interfering with CD44/ErbB-dependent signaling. *J Biol Chem* 2010;286(2):1475-85.
62. Zheng PS, Reis M, Sparling C, Lee DY, La Pierre DP, Wong CK, Deng Z, Kahai S, Wen J, Yang BB. Versican G3 domain promotes blood coagulation through suppressing the activity of tissue factor pathway inhibitor-1. *J Biol Chem* 2006;281(12):8175-82.
63. Schonherr E, Kinsella MG, Wight TN. Genistein selectively inhibits platelet-derived growth factor-stimulated versican biosynthesis in monkey arterial smooth muscle cells. *Arch Biochem Biophys* 1997;339(2):353-61.
64. Cardoso LE, Little PJ, Ballinger ML, Chan CK, Braun KR, Potter-Perigo S, Bornfeldt KE, Kinsella MG, Wight TN. Platelet-derived growth factor differentially regulates the expression and post-translational modification of versican by arterial smooth muscle cells through distinct protein kinase C and extracellular signal-regulated kinase pathways. *J Biol Chem* 2010;285(10):6987-95.
65. Schonherr E, Jarvelainen HT, Sandell LJ, Wight TN. Effects of platelet-derived growth factor and transforming growth factor-beta 1 on the synthesis of a large versican-like chondroitin sulfate proteoglycan by arterial smooth muscle cells. *J Biol Chem* 1991; 266(26): 17640-17647.
66. Evanko, S.P., Angello, J.C., and Wight, T.N. Formation of hyaluronan- and versican-rich pericellular matrix is required for proliferation and migration of vascular smooth muscle cells. *Arterioscler Thromb Vasc Biol* 1999;19(4):1004-13.
67. Evanko SP, Johnson PY, Braun KR, Underhill CB, Dudhia J, Wight TN. Platelet-derived growth factor stimulates the formation of versican-hyaluronan aggregates and pericellular matrix expansion in arterial smooth muscle cells. *Arch Biochem Biophys* 2001;394(1):29-38.
68. Zhang Y, Cao L, Yang BL, Yang BB. The G3 domain of versican enhances cell proliferation via epidermal growth factor-like motifs. *J Biol Chem* 1998; 273(33):21342-51.

69. Wu Y, Zhang Y, Cao L, Chen L, Lee V, Zheng PS, Kiani C, Adams ME, Ang LC, Paiwand F, Yang BB. Identification of the motif in versican G3 domain that plays a dominant-negative effect on astrocytoma cell proliferation through inhibiting versican secretion and binding. *J Biol Chem* 2001; 276(17): 14178-86.
70. Zhang Y, Cao L, Kiani C, Yang BL, Hu W, Yang BB. Promotion of chondrocyte proliferation by versican mediated by G1 domain and EGF-like motifs. *J Cell Biochem* 1999;73(4):445-57.
71. Perissinotto D, Iacopetti P, Bellina I, Doliana R, Colombatti A, Pettway Z, Bronner-Fraser M, Shinomura T, Kimata K, Morgelin M, Lofberg J, Perris R. Avian neural crest cell migration is diversely regulated by the two major hyaluronan-binding proteoglycans PG-M/versican and aggrecan. *Development* 2000;127(13):2823-42.
72. Perris R, Perissinotto D, Pettway Z, Bronner-Fraser M, Morgelin M, Kimata K. Inhibitory effects of PG-H/aggrecan and PG-M/versican on avian neural crest cell migration. *FASEB J* 1996;10(2):293-301.
73. Dutt S, Kleber M, Matasci M, Sommer L, Zimmermann DR. Versican V0 and V1 guide migratory neural crest cells. *J Biol Chem* 2006;281(17):12123-31.
74. Fidler PS, Schuette K, Asher RA, Dobbertin A, Thornton SR, Calle-Patino Y, Muir E, Levine JM, Geller HM, Rogers JH, Faissner A, Fawcett JW. Comparing astrocytic cell lines that are inhibitory or permissive for axon growth: the major axon-inhibitory proteoglycan is NG2. *J Neurosci* 1999;19(20):8778-88.
75. Jones LL, Sajed D, Tuszynski MH. Axonal regeneration through regions of chondroitin sulfate proteoglycan deposition after spinal cord injury: a balance of permissiveness and inhibition. *J Neurosci* 2003; 23(28): 9276-9288.
76. Tang X., Davies, J.E., and Davies, S.J. Changes in distribution, cell associations, and protein expression levels of NG2, neurocan, phosphacan, brevican, versican V2, and tenascin-C during acute to chronic maturation of spinal cord scar tissue. *J Neurosci Res* 2003;71(13):427-44.
77. Asher RA, Morgenstern DA, Shearer MC, Adcock KH, Pesheva P, Fawcett JW. Versican is upregulated in CNS injury and is a product of oligodendrocyte lineage cells. *J Neurosci* 2002;22(6):2225-36.
78. Yamada KM, Even-Ram S. Integrin regulation of growth factor receptors. *Nat Cell Biol* 2002;4(4):E75-6.
79. Adelsman MA, McCarthy JB, Shimizu Y. Stimulation of beta1-integrin function by epidermal growth factor and heregulin-beta has distinct requirements for erbB2 but a similar dependence on phosphoinositide 3-OH kinase. *Mol Biol Cell* 1999; 10(9): 2861-78.
80. Cabodi S, Moro L, Bergatto E, Boeri Erba E, Di Stefano P, Turco E, Tarone G, Defilippi P. Integrin regulation of epidermal growth factor (EGF) receptor and of EGF-dependent responses. *Biochem Soc Trans* 2004;32(Pt 3):438-42.
81. Lee JY, Spicer AP. Hyaluronan: a multifunctional, megaDalton, stealth molecule. *Curr Opin Cell Biol* 2000; 12(5): 581-6.
82. Henderson DJ, Copp AJ. Versican expression is associated with chamber specification, septation, and valvulogenesis in the developing mouse heart. *Circ Res* 1998;83(5):523-32.
83. Kern CB, Twal WO, Mjaatvedt CH, Fairey SE, Toole BP, Iruela-Arispe ML, Argraves WS. Proteolytic cleavage of versican during cardiac cushion morphogenesis. *Dev Dyn* 2006;235(8):2238-47.
84. Yamagata M, Yamada KM, Yoneda M, Suzuki S, Kimata K. Chondroitin sulfate proteoglycan (PG-M-like proteoglycan) is involved in the binding of hyaluronic acid to cellular fibronectin. *J Biol Chem* 1986;261(29):13526-35.
85. Wijelath ES, Murray J, Rahman S, Patel Y, Ishida A, Strand K, Aziz S, Cardona C, Hammond WP, Savidge GF, Rafii S, Sobel M. Novel vascular endothelial growth factor binding domains of fibronectin enhance vascular endothelial growth factor biological activity. *Circ Res* 2002;91(1):25-31.
86. LeBaron RG, Zimmermann DR, Ruoslahti E. Hyaluronate binding properties of versican. *J Biol Chem* 1992;267(14):10003-10.
87. Hasegawa K, Yoneda M, Kuwabara H, Miyaishi O, Itano N, Ohno A, Zako M, Isogai Z. Versican, a major hyaluronan-binding component in the dermis, loses its hyaluronan-binding ability in solar elastosis. *J Invest Dermatol* 2007;127(7):1657-63.
88. Mukaratirwa S, van Ederen AM, Gruys E, Nederbragt H. Versican and hyaluronan expression in canine colonic adenomas and carcinomas: relation to malignancy and depth of tumour invasion. *J Comp Pathol* 2004;131(4):259-70.
89. Suwihat S, Ricciardelli C, Tammi R, Tammi M, Auvinen P, Kosma VM, LeBaron RG, Raymond WA, Tilley WD, Horsfall DJ. Expression of extracellular matrix components versican, chondroitin sulfate, tenascin, and hyaluronan, and their association with disease outcome in node-negative breast cancer. *Clin Cancer Res* 2004;10(7):2491-8.

Versican; structure, function and regulation

90. Syrokou A, Tzanakakis GN, Hjerpe A, Karamanos NK. Proteoglycans in human malignant mesothelioma. Stimulation of their synthesis induced by epidermal, insulin and platelet-derived growth factors involves receptors with tyrosine kinase activity. *Biochimie* 1999;81(7):733-44.
91. Lemire JM, Merrilees MJ, Braun KR, Wight TN. Overexpression of the V3 variant of versican alters arterial smooth muscle cell adhesion, migration, and proliferation in vitro. *J Cell Physiol* 2002;190(1):38-45.
92. Harwood AJ. Regulation of GSK-3: a cellular multiprocessor. *Cell* 2001;105(7):821-4.
93. Deguchi JO, Yamazaki H, Aikawa E, Aikawa M. Chronic hypoxia activates the Akt and beta-catenin pathways in human macrophages. *Arterioscler Thromb Vasc Biol* 2009; 29(10): 1664-70.
94. Zhang X, Gaspard JP, Chung DC. Regulation of vascular endothelial growth factor by the Wnt and K-ras pathways in colonic neoplasia. *Cancer Res* 2001;61(16):6050-4.
95. Yoon H, Liyanarachchi S, Wright FA, Davuluri R, Lockman JC, de la Chapelle A, Pellegata NS. Gene expression profiling of isogenic cells with different TP53 gene dosage reveals numerous genes that are affected by TP53 dosage and identifies CSPG2 as a direct target of p53. *Proc Natl Acad Sci USA* 2002;99(24):15632-7.
96. Harris CC. Structure and function of the p53 tumor suppressor gene: clues for rational cancer therapeutic strategies. *J Natl Cancer Inst* 1996; 88(20): 1442-55.
97. Bossi G, Sacchi A. Restoration of wild-type p53 function in human cancer: relevance for tumor therapy. *Head Neck* 2007;29(3):272-84.