

The Potential of Circulating Tumor Cells in Personalized Management of Breast Cancer: A Systematic Review

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Abstract- Circulating tumor cells (CTCs) recognition and characterization in the peripheral blood of patients with breast cancer have proven practical and predictive value in different studies. However, the clinical significance of CTCs enumeration and molecular characterization in the personalization of breast cancer diagnosis and treatment remains under the debate. A literature search in PubMed, Web of Science and Scopus was performed from October 1990 to June 2016 for studies which evaluating CTCs and its association with clinical and pathological characteristics and medical outcome in the field of breast cancer personalization for both diagnosis and treatment categories. The treatment outcomes were progression-free survival (PFS) and overall survival (OS) or relapse in different patients. Sixty-nine studies met the inclusion criteria. The sample size varies from 1 to 2026. Median follow-up was 15 months (range 3-27). Different molecular techniques have been applied to research, but they mostly are based on CTCs enrichment and then detection by using FDA-approved Cell SearchTM. By far the most studies define CTCs as cytokeratins (CK) positive and CD45 negative cells. Despite the differences in methodology, twenty-eight studies for breast cancer diagnosis and prognosis were mainly focused on CTCs isolation and enumeration. Forty-three researches were about CTCs count and exact molecular characterization. In the way of precision treatment, CTCs detection before starting the first-line of therapy or during therapy in breast cancer patients is extremely valuable, but in the way of precision medicine it should be supported with some molecular characteristics of CTCs like CTCs phenotypic changes, gene expression analysis of CTCs and molecular characteristics of CTCs.

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Introduction

Breast cancer is the most common type of cancer amongst women in both developed and developing countries (1). According to American Cancer Society, the new breast cancer cases among women in 2012 was 1,676,600. The number of breast cancer deaths in women in 2012 was 521,900 all over the world (2). Most women undergo surgery for breast cancer and also

receive other treatment such as hormone therapy, chemotherapy or radiation before or after surgery (lumpectomy, mastectomy, sentinel node biopsy, auxiliary lymph node dissection or removing both breasts). One of the problematic issues about breast cancer is drug resistance and tumor relapse which occurred in an unpredictable level in different patients that means not all patients respond equally to cancer therapeutic compounds. At the molecular level, how a

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Circulating tumor cells

person responds to a cancer therapy is running in their gene expression pattern, genetic changes and their position in the cancer genome (3-5). The reference book from the WHO clusters breast cancer into 17 different types according to their microscopic appearance (6) and The genomic and transcriptomic architecture of 2,000 breast tumors discloses novel subgroups (7).

Thanks to the use of biotechnologies, impressive steps toward understanding the biology of breast cancer have been accomplished over last decade. In order to discover the genomic characteristics of breast cancer, new generation of biomarkers has become available with the discovery of the genetic alterations that are responsible for the initiation and progression of human breast cancers (8-10). Because of breast cancer intra-tumor heterogeneity (11) for real-time monitoring of the treatment, there is an essential need to repeat tumor biopsies from different anatomical areas and at different time-points. However, common tissue biopsies of a small tumor region may not provide an exact characterization of the genetic, epigenetic and/or phenotypic alterations found in the tumor as a whole (12,13). Additionally, it is quite challenging since it is costly, painful, hard to repeat and potentially risky for the patient.

For cancer research, liquid biopsies, which are a diagnostic concept, open a new perspective for real time tracking of cancer. Liquid biopsy is defined as circulating tumor cells (CTCs) and fragments of tumor DNA (ctDNA) that are released into the blood from the primary tumor and from metastatic sites (14). As we know for tumor metastasis the spread of a primary tumor to the blood stream through CTCs is a critical step (15). Circulating tumor cells (CTCs) are cancerous cells originating from a primary or metastatic tumor and shed into the peripheral blood (16). In breast cancer (BC), CTCs are detectable in patients with both early stages and late stages of disease (17-19). It has been shown that the CTCs detection may help to predict the clinical outcome in patients with different types of cancers, especially the enumeration of CTCs before starting systemic treatment in both metastatic and non-metastatic breast cancer patients (20). Furthermore, CTC count at different time points during systemic treatment could be a reliable marker of treatment response and have to decide on therapies based on molecular characteristics of CTCs (21-24). Because CTCs are found in circulation as a collectible fraction that is representative of the tumor, they may provide an ideal model to study the biology of the tumor at various intervals before and during treatment (23,24).

Take everything into consideration; precision breast

cancer treatment can be possible by using CTCs enumeration or characterization (25). Interestingly, several authors have shown that monitoring CTC levels facilitate prediction of treatment efficacy (26,27). In this article, we provide a first-time systematic review about research focusing on using both CTCs enumeration and molecular characterization and personalization of therapeutic and diagnostic procedures of breast cancer.

Materials and Methods

An independent systematic review of the literature across PubMed, web of Science and Scopus was conducted in July 2016. The search strategy included keywords such as “CTCs” or “Circulating Tumor Cells” or “liquid biopsy” and “breast cancer” and “personalized medicine” or “precision therapy” or “P4 medicine” or “stratified medicine” through their title, abstract and text from October 1990 to June 2016. Only studies published in peer-reviewed journals were included, data from letters and conference abstracts or report were not included. The study selection process is shown in Figure 1 and search strategies, and results are provided in additional File 1. Two reviewers evaluated all the candidate titles and abstracts categorized by the search strategy, and all potentially relevant publications were retrieved in full. They independently evaluated the selected articles for study eligibility. After a preliminary review of articles for study inclusion, an inter-reviewer agreement was assessed with the Cohen's kappa (κ) coefficient, and disagreement was resolved by discussion (16).

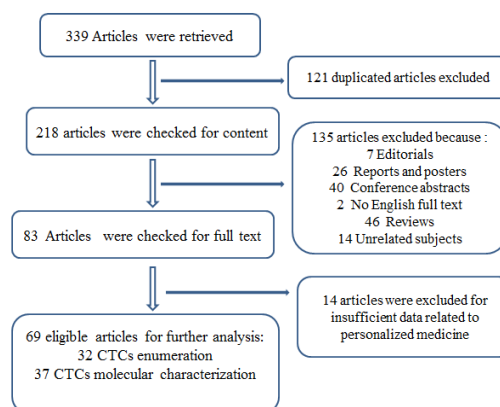


Figure 1. PRISMA flow diagram presenting the results of the literature search and study selection process

Data extraction

For eligible studies specific data elements included the following: author, year of publication, journal citation,

Continuance of Table 1.

De Giorgi, U.(30)	Circulating tumor cells and bone metastases as detected by FDG-PET/CT in patients with metastatic breast cancer	2010	Case series	195	MBC	CTCs enumeration	FDG-PET/CT, CellSpector Analyzer (Immuncor Corporation, Huntington PA)	The CellSearch System (Verтек, Immuncor), CellSpector Analyzer (Immuncor), Iliad/Idexys/Genos (Iliad/Idexys/Genos), Iliad/Idexys/Genos (Iliad/Idexys/Genos), Iliad/Idexys/Genos (Iliad/Idexys/Genos)	lacking CD45 and expressing cytokeratin	CTCs : nucleated cells that lacked CD45 and expressed cytokeratin	Adjuvant hormonal therapy Hormonal therapy HER2 target therapy	Systemic therapy	HER2 target therapy	Presence of extensive bone metastases as detected by FDG-PET/CT is associated with increased CTC numbers in MBC	Detection of five or more CTCs during therapeutic monitoring can accurately predict prognosis in MBC beyond metabolic response. FDG-PET/CT deserves a role in patients who have fewer than five CTCs at midtherapy.
De Giorgi, U.(38)	Circulating tumor cells (CTC), Iliad/Idexys/Genos (Iliad/Idexys/Genos), Iliad/Idexys/Genos (Iliad/Idexys/Genos), Iliad/Idexys/Genos (Iliad/Idexys/Genos)	2009	Case series	115	MBC	Comparison the prognostic value of CTC and FDG-PET/CT	The CellSearch System (Verтек, Immuncor), CellSpector Analyzer (Immuncor), Iliad/Idexys/Genos (Iliad/Idexys/Genos), Iliad/Idexys/Genos (Iliad/Idexys/Genos), Iliad/Idexys/Genos (Iliad/Idexys/Genos)	CTCs enumeration	CTCs : nucleated cells that lacked CD45 and expressed cytokeratin	epithelial adhesion molecules (EPCAM)	Adjuvant systemic treatment (hormonal therapy or chemotherapy)	Systemic therapy	CTCs enumeration	Presence of CTC in breast cancer patients before undergoing surgery with curative intent is associated with an increased risk for breast cancer-related death	Presence of CTC in breast cancer patients before undergoing surgery with curative intent is associated with an increased risk for breast cancer-related death
Franken, Bast(90)	Circulating tumor disease recurrence and survival in newly diagnosed breast cancer	2012	case series	602	Breast Cancer	CTCs enumeration	The CellSearch System (Verтек, Immuncor), Iliad/Idexys/Genos (Iliad/Idexys/Genos), Iliad/Idexys/Genos (Iliad/Idexys/Genos)	CTCs enumeration	epithelial adhesion molecules	cell	Adjuvant systemic treatment (hormonal therapy or chemotherapy)	Systemic therapy	CTCs enumeration	Presence of CTC in breast cancer patients before undergoing surgery with curative intent is associated with an increased risk for breast cancer-related death	Presence of CTC in breast cancer patients before undergoing surgery with curative intent is associated with an increased risk for breast cancer-related death
Giordano, A.(45)	Artificial neural network analysis of circulating tumor cells in metastatic breast cancer patients	2011	Cases series	311	MBC	CTCs enumeration	artificial neural network (ANN)	CTCs enumeration	ER, PR, HER2	Chemotherapy Hormonal therapy Anti-HER2 drug	Chemotherapy Hormonal therapy Anti-HER2 drug	Neural network analysis	Neural network analysis	Neural network analysis	Neural network analysis
Giuliano, M(43)	Circulating tumor cells as prognostic and predictive markers in breast cancer patients receiving first-line therapy	2010	Case series	235	MBC	CTCs enumeration	CTC assessment performed with CellSearch	CTCs enumeration	epidermal growth factor receptor-2	epithelial cell adhesion molecule (EPCAM)-positive, cytokeratin (CK)-positive, DAPI-positive, and CD45-negative	Tasuzumab Lapatinib	Radiation Chemotherapy	Prognostic information provided by CTC count may be useful in patient stratifications and therapeutic selection (particularly in the group with positive CTCs)	Prognostic information provided by CTC count may be useful in patient stratifications and therapeutic selection (particularly in the group with positive CTCs)	
Green, T. L.(31)	Circulating tumor cells from metastatic breast cancer patients linked to decreased immune function and response to treatment	2013	Case series	45	MBC	CTC enumeration and NK cell function measurement	The CellSearch Cell Kit (Verтек, South Plainfield, NJ), Radionuclide assay	CTC enumeration and NK cell function measurement	epithelial cell adhesion molecule (EPCAM)-positive, cytokeratin (CK)-positive, DAPI-positive, and CD45-negative	epithelial cell adhesion molecule (EPCAM)-positive, cytokeratin (CK)-positive, DAPI-positive, and CD45-negative	Radiation Chemotherapy	Radiation Chemotherapy	CTC enumeration and NK cell function measurement	CTC enumeration and NK cell function measurement	CTC enumeration and NK cell function measurement
Hall, C.(39)	Circulating tumor cells after Neoadjuvant Therapy in Stage I-II Triple-Negative Breast Cancer	2015	Case series	57	TNBC(triple negative breast cancer)	CTCs enumeration	Cell Search System (Amersco), A serotonaminated fluorescence-based microscope system	CTCs enumeration	cells positive for CK and negative for CD45	cells positive for CK and negative for CD45	Neoadjuvant chemotherapy (NACT)	Neoadjuvant chemotherapy (NACT)	CTC enumeration	CTC presence was not associated with primary tumor size, high grade, or lymph node positivity, or more CTCs present after NACT mediated relapse and survival in nonmetastatic TNBC patients.	CTC presence was not associated with primary tumor size, high grade, or lymph node positivity, or more CTCs present after NACT mediated relapse and survival in nonmetastatic TNBC patients.
Hall, C. S.(41)	Circulating Tumor Cells and Recurrence After Primary Systemic Therapy in Stage III Inflammatory Breast Cancer	2015	case series	63	stage III Inflammatory breast cancer (IBC)	CTCs enumeration	The CellSearch System (Amersco)	CTCs enumeration	lacking CD45 but expressing cytokeratins (CK) 8, 18, or 19	lacking CD45 but expressing cytokeratins (CK) 8, 18, or 19	Primary systemic chemotherapy	Primary systemic chemotherapy	CTCs enumeration	CTCs after primary chemotherapy identified patients at high risk for relapse	CTCs after primary chemotherapy identified patients at high risk for relapse
Hartkopf, A. D.(58)	Changing levels of circulating tumor cells in monitoring chemotherapy response: patients with metastatic breast cancer	2011	Case series	58	Advanced MBC	CTCs and CA 15-3 enumeration	CellSaver urecs (Verтек, Warren, NJ, USA), sequential chemiluminescent sandwich immunoassay on the ADVIA Centaur System (Siemens Diagnostics Eschborn, Germany)	CTCs and CA 15-3 enumeration	serum CA 15-3 measurement	serum CA 15-3 measurement	Chemotherapy, in combination with targeted therapy.	Chemotherapy, in combination with targeted therapy.	CTCs and CA 15-3 enumeration	Changing CTC levels during chemotherapy are useful to monitor therapy efficacy.	Changing CTC levels during chemotherapy are useful to monitor therapy efficacy.
Hayes, D. F.(120)	Circulating tumor cells at each follow-up time point during therapy of metastatic breast cancer patients predict progression-free and overall survival	2006	Clinical Trial (case series)	177	MBC	CTCs enumeration	CellSearch System (Verтек, Immuncor), CellSpector Analyzer (Immuncor)	CTCs enumeration	nucleated cells lacking CD45 and expressing cytokeratin	nucleated cells lacking CD45 and expressing cytokeratin	Systemic therapy	Systemic therapy	CTCs enumeration	Detection of elevated CTCs at any time during therapy is an accurate indication of subsequent rapid disease progression and mortality for MBC patients	Detection of elevated CTCs at any time during therapy is an accurate indication of subsequent rapid disease progression and mortality for MBC patients
Karhade, M.(40)	Circulating tumor cells in non-metastatic triple-negative breast cancer	2014	case series	113	stages I-III TNBC	CTCs enumeration	CellSearch System (Verтек, Immuncor), CellSpector Analyzer (Immuncor)	CTCs enumeration	epithelial-cell-adhesion molecule, lacking CD45 but expressing cytokeratins (CK)	epithelial-cell-adhesion molecule, lacking CD45 but expressing cytokeratins (CK)	Neoadjuvant chemotherapy	Neoadjuvant chemotherapy	CTCs enumeration	Two or more CTCs predict shorter progression-free and overall survival in TNBC patients	Two or more CTCs predict shorter progression-free and overall survival in TNBC patients

Continuance of Table 1.

Lin, M. C.(12)	Lucci, Anthony(122)	Mizzi, M.(65)	Müller V(42)	Nakamura, S.(123)	Note, F.(92)	Pechmann, K.(124)	Peters, D. J.(46)	Perra, J. Y.(36)	Perra, J.-Y.(35)
Circulating Tumor Cells: A Useful Predictor of Treatment Efficacy in Metastatic Breast Cancer	Circulating tumor cells in non-metastatic breast cancer: a prospective study	Frequent expression of PD-L1 on circulating breast cancer cells	Prognostic impact of circulating tumor cells assessed with the CellSearch System™ and AdnaTest Breast in metastatic breast cancer patients: the DETECT study	Multi-center study evaluating circulating tumor cells as a surrogate for response to treatment and overall survival in metastatic breast cancer	Variation of circulating tumor cell levels during treatment of metastatic breast cancer: prognostic and therapeutic implications	Monitoring the response of epithelial tumor cells to adjuvant chemotherapy in breast cancer patients at risk of early relapse	Detection and prognostic significance of circulating tumor cells in patients with metastatic breast cancer according to Zimmunistochemical subtypes	Circulating tumor cell detection predicts early relapse after neoadjuvant chemotherapy in large operable and locally advanced breast cancer in a phase II randomized trial	High independent prognostic value of circulating tumor cells compared with serum tumor markers in a large prospective trial in first-line chemotherapy for metastatic breast cancer
2009	2012	2015	2012	2010	2008	2008	2014	2008	2012
Case series	Case-Control	Case-Control	Case series	Case series	Case series	Case series	Case series	Case series	Case series
68+ 6	302.(73-229)	31(1615)	254	119	80	91	154	118	267
MBC	stage 1-3 breast cancer undergoing surgery	MBC	MBC	MBC	advanced breast cancer patients	nonmetastatic primary breast cancer patients	Luminal B-HER2- negative, Luminal B-HER2-positive +HER2- positive (non-luminal) Triple negative, Not	MBC	MBC
Enumeration of CTCs and Definition of Response by Radiographic Imaging	CTCs enumeration	CTC enumeration (was used for these specific experiments)	CTCs enumeration	CTCs enumeration	CTCs enumeration	Analysis of circulating epithelial tumor cells (CTCs)	CTCs enumeration	CTCs enumeration	CTCs enumeration and comparison with serum tumor markers (CA 15-3, carcinoembryonic antigen and lactate dehydrogenase)
CellSearch System (Veridex, Raritan, NJ), semiautomated/fluorescence-based microscope system	Western blot, Flow cytometry and immunocytochemical analyses (CellSearch, AdnaTest, Cytocell, CellSearch Kit (Janssen)	CellSearch System (Veridex, Raritan, NJ), semiautomated/fluorescence-based microscope system	AdnaTest Breast Cancer and the CellSearch System	Cell Traks Analyzer (Veridex LLC, Raritan, NJ), CellSearch System (Veridex, Raritan, NJ), CellSearch™ Epithelial Cell Kit, semiautomated fluorescence microscopy system	CellSearch System (Veridex) (CellTrack-AutoPrep system-CellSpatier Analyzer	Panometricgraphs of epithelial cells with green fluorescent antibody (CellSearch system (Veridex, Warren, NJ)	FISH ,IHC ,CellSearch system (Veridex, LLC) using the IVD CellSearch CTC kit	The standardized CellSearch technique, Semiautomated fluorescence-based microscopy system	CTCs were counted with CellSearch®
HER2/neu results, cytokeratin positive, DAPI positive, and CD45 negative	metastatic cells for cytokeratin and negative for CD45	PD-L1(CD279), PD-L2 (B7-DC, CD273), EpCAM-AMPH-CK+ CD45	HER2, MUC1, and GAT3-2, EpCAM, DAPI staining	CTCs: medeted cells lacking CD45 and expressing cytokeratin	CTCs:coimmunohlogor ya DAPI positive, positivity for cytokeratin and negative staining for CD45	ER expression	EpCAM, (ER)PR, HER2/neu status	CD45-allelophycocyanin and epithelial cells (cyokeratin 8,18,19-phycoerythrin	CTC count, CEA, CA 15-3, LDH and ALP
Chemotherapy, +biologic agent Endocrine therapy + biologic agent Biologic agent alone	Adjuvant chemotherapy	Anti-PDL1 antibody therapy	Anti HER2 target therapy	Chemo+ trastuzumab, Chemo + trastuzumab + Hormone + Hormone + Trastuzumab	With a maximum two lines of therapy.	Adjuvant chemotherapy	Cytotoxic Endocrine treatment (including trastuzumab, lapatinib, pertuzumab and TDM1) and bisphosphonates alone.	Preneoadjuvant chemotherapy and/or postneoadjuvant chemotherapy	Chemotherapy with/without targeted therapy(taxanes , anthracyclines , 5-fluorouracil and vinorelbine
A strong correlation between CTC results and radiographic disease progression in patients receiving chemotherapy or endocrine therapy for MBC.	The presence of one or more circulating tumor cells predicted early recurrence and decreased overall survival in chemotherapy patients with non-metastatic breast cancer	PD-L1 is frequently expressed on metastatic cells circulating in CTC/PP-1 assay can be used for liquid biopsy in future clinical trials for stratification and monitoring of cancer patients undergoing immune checkpoint blockade.	The present results indicate that the CellSearch system is superior to AdnaTest Breast Cancer in predicting clinical outcome in breast cancer	Because the change in the number of CTCs was highly correlated with results from imaging before and after therapy, CTCs can be considered a biomarker that may predict the effect of treatment earlier than imaging modalities.	CTCs basal value is a predictive indicator of prognosis and changes in CTC levels during therapy may indicate a clinical response. Testing CTC levels targeted treatments might substitute measurement parameters for response evaluation.	Peripherally circulating tumor cells are influenced by systemic chemotherapy and that an increase (even after initial response) to therapy of 10-fold or more at the end of therapy is a strong predictor of relapse and a surrogate marker for aggressiveness of the tumor cells	The detection of EpCAM/CTCs was not clearly associated with any of the immunohistochemical subtypes of breast cancer in patients with MBC before first-line treatment. Potentially clinically relevant differences were however observed at very high CTC counts. Furthermore, our data suggest a lower prognostic significance of CTC evaluation in HER2-positive patients with MBC.	Circulating tumor cells can be detected by the CellSearch system at a low cutoff of 1 cell in 27% of patients receiving neoadjuvant chemotherapy. Circulating tumor cell detection was not correlated to the primary tumor response but is an independent prognostic factor for early relapse	This is the largest prospective validating study of the prognostic value of CTC independently from serum tumor marker. Elevated CTCs before C2 are an early predictive marker of poor PFS and OS which could be used to monitor treatment benefit. CTC decrease under treatment seems stronger with targeted therapy.

Continuance of Table 1.

Rack, Brigitte(125)	Siedt, A. (126)	Serrano, M. J.(127) Serrano, M. J.(67)	Wallwiener, M.(93)
Circulating Tumor Cells Predict Survival in Early Average-to-High Risk Breast Cancer Patients	Circulating tumor cell enumeration with a combination of epithelial cell adhesion molecule- and cell-surface vimentin-based methods for monitoring breast cancer therapeutic response	Detection of circulating tumor cells in the context of treatment Prognostic value in breast cancer Dynamics of circulating tumor cells in early breast cancer under neoadjuvant therapy	Serial enumeration of circulating tumor cells predicts treatment response and prognosis in metastatic breast cancer: a prospective study in 393 patients
2014	2015	2009 2012	2014
Cohort	Case series	Case series Case series	Case series
2026 / 1492	58	92(71) 26	393
early breast cancer	MBC	Breast cancer (metastatic and non metastatic) non-metastatic breast cancer patients	MBC
CTCs enumeration	CTCs enumeration	CTCs enumeration CTCs enumeration	CTC status at baseline (CTCBL) and after one cycle of a new line of systemic therapy (CTCIC) and CTCIN enumeration
Immuno-magnetic enrichment for cells expressing the epithelial-cell adhesion molecule (CellSearch System (Veridex, Raritan, NJ) CellSearch System	CellSearch system, enumeration of CTCs by Her1 and/or E-cadherin, Her2 and CD5, Dependent Kit (Stem Cell Technologies), Nuclein EpCAM Staining	N/A Flow gradient and selective immunomagnetic cell separation, immunocytochemical staining	CellSearch™ assay (CellSearch System) Epithelial Cell Adhesion Molecule (ECAM) Staining Veridex LLC, Raritan, NJ, USA)
CTCs were defined as nucleated cells expressing cytokeratin and lacking CD45	EpCAM, 84-1 and CD45 and epithelial/mesenchymal ratio	N/A cytokeratin 7, 8, 18 and 19	Cytokeratin (CK) 8, 18, and 19, and lacking CD45
Primary operation: Breast conserving, Mastectomy Radiotherapy: Systemic: Performed/Not performed Chemotherapy/Chemotherapy/Endocrine treatment Trazosunab	-	Chemotherapy Neoadjuvant treatment	Systemic treatment
These results suggest the independent prognostic relevance of CTCs both before and after adjuvant chemotherapy in a large prospective trial of patients with primary breast cancer.	The specificity of CTC detection was found to be highest when the sum of CTC counts from the 2 methods was above a threshold of 8 CTCs/7.5 ml. The sum of CTC counts from the CellSearch and CSV methods appears to provide new insights for assessment of therapeutic response and this provides a new approach to personalized medicine in breast cancer patients.	The differential prognostic and overall survival showed between patients with and without elevated CTCs before and at the end of chemotherapy, is of special interest in clinical evidence of metastasis. Molecular and genetic characterization of CTCs, chemoresistance profiles should also be able to advise the clinician regarding the most efficacious chemotherapy regimens. In terms of tumor biology, it is clear that circulating tumor cells are present in early breast cancer, thus supporting the theory of early metastasis	CTCBL, CTCIC, and CTCIN are predictive of outcome in MBC. Serial CTC enumeration is useful in tailoring systemic treatment of MBC

They check CTCs number at the starting point, through the first weeks of treatment and after treatment completion as a factor for progression-free survival (PFS), overall survival (OS) and relapse in different patients. CTCs are defined mostly in place of cytokeratin (CK) positive and CD45 negative cells. Detection of five or more CTCs per 7.5 mL blood during therapeutic monitoring can accurately predict prognosis in MBC (30) and significantly decreased responses by their immune cells in comparison with those patients who had 5 CTCs or less (20,31,32) so it is a strong prognostic factor for OS during neoadjuvant chemotherapy (NACT) in MBC patients (33-36). About positron emission tomography-computed tomography (PET) it can easily say that FDG-PET/CT and FLT-PET and CTC analyses could be considered to potentially predict early response when used in combination; correlations with OS and PFS (37,38). One or more CTCs present after neoadjuvant chemotherapy predicted

relapse and survival in non-metastatic triple-negative breast cancer (TNBC) patients but CTCs presence was not connected to the primary tumor size, high grade or lymph node positivity (39,40) and also CTCs after primary chemotherapy recognized inflammatory breast cancer (IBC) patients who are at risk of relapse (41). The results indicate that the CellSearch™ system is superior to the DNA Test in the way of clinical outcome in advanced breast cancer prediction (42). Finally, prognostic information provided by CTC count may be useful in patient stratifications and therapeutic selection (particularly in the group with positive CTCs) (43), but CTCs were powerfully predictive of survival in all MBC subtypes excluding Her2 positive patients who had been received targeted therapy (44,45). Some data propose a lower prognostic implication of CTC evaluation in Her2-positive patients with MBC (46).

Additional thirty-seven researches mostly consider cellular markers and gene expression profile of CTCs

and have more emphasize on personalized breast cancer diagnosis and treatment (Table 2). In twenty-one of them the most common molecular marker was a proto-oncogene Neu (Her2) alone or collectively with other molecules such as epithelial cell adhesion

molecule (EPCAM), progesterone receptor (PR, also known as NR3C3 or nuclear receptors subfamily 3, group C, member 3) and estrogen receptors (ERs) (47).

Table 2. Using of CTCs molecular characteristics in personalized management of Breast Cancer

First Author	Name of study	Year	Patients number	Type of study	Type of breast cancer	Evaluation targets	Methods	Personalized target molecules	Type of treatment	Conclusion
Ageliki, S.(5)	Efficacy of Lapatinib in Therapy-Resistant Circulating Tumor Cells in Metastatic Breast Cancer	2015	22	Clinical Trial (case series)	MBC	HER2 positive CTCs count	Immunofluorescent microscopy	EGFR, HER2	Lapatinib	Lapatinib is effective in decreasing HER2-positive CTCs in patients with MBC irrespectively of the HER2 status of the primary tumor
Apolski, S.(50)	Use of circulating tumor cells in the peripheral blood of patients with stage I and II breast cancer after the administration of adjuvant chemotherapy: evaluation of their	2007	214	Case series	BC (Stage I and II)	HER2 mRNA-positive CTCs detection	Nested RT-PCR	HER2 mRNA	Adjuvant chemotherapy	positive CTCs are the completion of the adjuvant chemotherapy may provide clinically useful information concerning the efficacy of treatment
Ascolini, G.(29)	Modelling Circulating Tumor Cells for Personalized Survival Prediction in Metastatic Breast Cancer	2015	N/A	Case series	The earliest stage of BC	CTCs gene expression	Branching process model	ERCAM, CD47, CD44 and MET	Bisphosphonates-therapy	metastatic breast cancer patients survival probability by modifying the populations of circulating tumour cells and it could be extended to other
de Albuquerque, A.(128)	Multimer Analysis of Circulating Tumor Cells in Peripheral Blood of Metastatic Breast Cancer Patients: A Step Forward in Personalized Medicine	2012	32 MBC /42 negative controls	Case-Control	MBC	Enumeration and characterization of CTCs	Cell Culture, Immunomagnetic Enrichment Antibodies, real-time reverse transcription-polymerase chain	BM7, VU109, KR19, SCGB2A, MUC1, EPCAM, BIRC5 and ERBB2	-	enumerating CTCs seems to be an important tool that might identify women who were initially ineligible for breast cancer therapy but who would later qualify for
De Luca, F.(55)	Mutational analysis of single circulating tumor cells by next generation sequencing in metastatic breast cancer	2016	4	Case series	Stage III and IV BC	Molecular characterization of single CTCs	Next Generation Sequencing (NGS)	50 cancer related genes	Chemotherapy, Endocrine therapy	CTC characteristics are more closely linked to the dynamic modifications of the disease status
Dvella, R.(71)	Use of circulating tumor factor-beta (TGF-beta) and fibronectin (FN) as predictors of distant seeding of circulating tumor cells in patients with metastatic breast	2013	65	Case series	non-treated stage III-IV MBC	Enumeration and characterization of CTCs	ELISA, ELISA, AdhT estBrestCancerTest (AdhGen AG, Langenlengen, Germany; method B)	CK3+, CTCS, Transforming growth factor-beta (TGF-beta) and Chemokine (C-X-C Motif) Ligand-1 (CXCL1)	Systemic therapy	TGF-β and CXCL1 are associated with a poor prognosis, and higher detection of CTCs and propensity of these cells to seed lung metastases in patients with breast
Fehm, T.(99)	HER2 status of circulating tumor cells in 15 patients with metastatic breast cancer: a prospective, multicenter trial	2010	254	Cohort	MBC	Enumeration and characterization of CTCs	CellSearch assay	HER2	HER2-targeted therapies	relevant number of patients with HER2 negative primary tumors. Therefore, it will be mandatory to correlate the assay-dependent HER2 status of CTCs
Fehm, T.(51)	Determination of HER2 status using both serum HER2 levels and circulating tumor cells in patients with recurrent breast cancer whose primary tumor was HER2 negative or of unknown HER2 status	2007	77 (44/33)	Case series	MBC	Evaluation of HER2 status of circulating tumor cells	Slide-based assay	GA 753-2, MUC1 or HER2	HER2 targeted therapy and endocrine therapy	patients with initially negative or unknown HER2 status can have elevated serum HER2 levels and/or HER2-positive CTCs at the time of development of
Fehm, T.(52)	Detection and characterization of circulating tumor cells in blood of primary breast cancer patients by RT-PCR and comparison to status of bone marrow disseminated cells	2009	431	Case series	primary breast cancer	correlation between CTCs and disseminated tumor cells (DTCs) in the bone marrow (BM) evaluation and CTCs Molecular characterization	AdhT estBrestCancerTest (AdhGen AG, Germany), RT-PCR, IHC	EPCAM, MUC1 and HER2 transcripts, Expression of the ER and PR	HER2 targeted therapy and endocrine therapy	the impact on adjuvant treatment can only be answered in clinical trials randomizing patients according to the expression profile based on CTCs or

Continuance of Table 2.

Hall C(99) Giordano, A.(44)	Gradione, A.(129)	Gradione, A.(130)	Igenatidis, M.(48)	Jansson, Sand(131)	Konig, A.(62)	Lipshart, S. T.(49)	Liu, Z(28)	Lowe, A. C.(94)
Circulating Tumor Cells after Neoadjuvant Therapy Predict Outcome in Stage I to III Breast Cancer Circulating tumor cells in immunohistochemical subtypes of metastatic breast cancer: lack of prediction in HER2-positive disease treated with targeted therapy	Circulating tumor cells (CTCs) in metastatic breast cancer (MBC): prognostic, drug resistance and phenotypic characterization	How Circulating Tumor Cells Escape From Multiple Resistance Mechanisms in Metastatic Breast Cancer Treatment	HER2-positive circulating tumor cells in breast cancer	Prognostic impact of circulating tumor cell apoptosis and clusters in serial blood samples from patients with metastatic breast cancer in a prospective observational cohort	Determination of Interleukin-4, -5, -6, -8 and -13 in Serum of Patients with Breast Cancer Before Treatment and its Correlation to Circulating Tumor Cells	Unbiased quantitative assessment of Her-2 expression of circulating tumor cells in patients with metastatic and non-metastatic breast cancer	Endoprotection of EGFR-positive circulating tumor cells and response with lapatinib and capcitabine	Young investigator challenge: Application of cytologic techniques to circulating tumor cell specimens: Detecting activation of the oncogene transcription factor STAT3
2011 2012	2011	2011	2011	2016	2016	2013	2010	2015
57 517	42	42	6 BC cell line	52	200	103 (M1) / 88 (M0)	1	52,42
case series Case series	Case series	Case series	Case/Control	Cohort	100 CTCs# / 100 CTCs	Case/Control	Case presentation	Case/Control
stage I to III Triple-negative breast cancer (TNBC) MBC	MBC	MBC	BC	All breast cancer subtypes	Breast Cancer	BC	MBC	BC
Emuneration and characterization of CTCs	CTCs isolation and molecular profiling	CTCs isolation and molecular profiling	CTCs enumeration and HER2 expression	Morphologic characterization of CTCs Emuneration and characterization of CTCs	Emuneration and characterization of CTCs	CTC enumeration and Her-2 assessment	Emuneration and characterization of CTCs	CTC enumeration and Characterization (noncancer patients was spiked with breast cancer cell lines)
fluorescence in situ hybridization (FISH) . CellSearch System (Janssen Diagnostics, LLC) Immunohistochemical (IHC) fluorescent in situ hybridization . CellSearch	PCR	CELLlectionDyna bead s coated with a monoclonal Antibodies	CellSearch, immunofluorescence	CellSearch System (Janssen Diagnostics), visual examination of the gillfilters, generated by the CellTrack-Analyzer I immunost microscope (CellTrack Analyzer v1)	CellSearch System (Janssen Diagnostics, South Raritan, NJ, USA, CellTracks Analyzer II (Janssen Diagnostics)	CellSearch(R) system, Her2-fluorescent immunocytochemical (IHC) fluorescence	CellSearch system, and FACTS analysis, IHC	Routine cytologic techniques, immunocytochemistry /immunohistochemistry
positive for CK and negative for CD45 HER2	MIRs, ALDH1, Etn and HER2/neu	multiple resistance-related proteins 1 and 2 (MRP1, MRP2)	HER2-positive CTCs	CTCs : CK+ /CD45- /DAPI+ cells fulfilling certain predefined criteria	Interleukin-4, -5, -6, -8 and -13/Th2 cytokines	Her-2- FTTc signal intensities	EGCAM positive but CD45 negative, estrogen receptor (ER) and progesterone receptor (PR) and were strongly positive for Her2	constitutive or inducible pSTAT3 expression and Ki-67
Neoadjuvant chemotherapy (NACT) predicted worse outcome in nonmetastatic TNBC patients HER2-targeted therapy + chemotherapy	New systemic therapy without limits to number of previous therapies	Antiherc/line-based chemotherapy, NPLD Nonpegylated liposomal doxorubicin (NPLD)	(Neo) adjuvant chemotherapy	First-line systemic therapy, Endocrine only Chemotherapy only HER2-directed (with trastuzumab) had worse prognosis than patients without these CTC characteristics. In patients with 55 CTC/7.5 ml blood at BL, morphologic characterization of	Randomized adjuvant therapy and endocrine therapy	Neoadjuvant chemotherapy	Neoadjuvant treatment, Trastuzumab, lapatinib-based treatments	Targeted therapy
One or more CTCs present after NACT predicted relapse and survival in nonmetastatic TNBC patients. CTCs were strongly predictive of survival in all MBC subtypes except HER2+ patients who had been treated with targeted therapy	The presence of CTCs expressing MIRs and ALDH1 is predictive of response to chemotherapy in MBC patients	who received conventional anthracyclines (doxorubicin or epirubicin), had a significantly shorter time to	were detected in IHCs, IHCs or M0 BC irrespective of the primary tumor HER2 status. Monitoring of HER2 expression on CTCs might be useful in trials with anti-	used for identifying true	In patients who are CTC-negative and progesterone receptor-positive, IL-4/Th2 cytokines are significantly modified	CTCs within each patient, it has the feasibility of unbiased quantitative and reproducible assessment of treatment targets on CTCs, opening a	associates with tumor response, whereas disease progression was related to a recurrence in CTCs, which were both EGFR and Her2 negative. Expression	immunogenetic analysis can be easily integrated into the existing clinical workflow, moving the field closer to a true peripheral blood liquid

Continuance of Table 2.

<p>Lu, J.(63) Mishonov, V.(66) Mesner B.(84)</p>	<p>Muller, V.(59)</p>	<p>Minzone, E.(53)</p>	<p>Nadler, R.(47)</p>	<p>Pestini, M.(113)</p>	<p>Polzer, B.(114)</p>	<p>Rajim, E. A.(56)</p>	<p>Reinholz, M. M.(132)</p>
<p>Isolation of circulating epithelial and tumor progenitor cells with an invasive phenotype from breast cancer patients Detection of circulating tumor cells during follow-up of patients with early breast cancer: Clinical utility for monitoring of therapy efficacy CD19f-based selection of circulating tumor cells (CTCs) improves detection across breast cancer subtypes.</p>	<p>Circulating tumor cells in breast cancer Correlation to bone marrow micrometastases, heterogeneous response to systemic therapy, and low proliferative activity</p>	<p>Changes of HER2 Status in Circulating Tumor Cells Compared With the Primary Tumor During Treatment for Advanced Breast Cancer</p>	<p>Biomarkers characterization of circulating tumour cells in breast cancer patients</p>	<p>Heterogeneity of PIK3CA mutational status at the single cell level in circulating tumor cells from metastatic breast cancer patients</p>	<p>Molecular profiling of single circulating tumor cells with diagnostic intention</p>	<p>An 8-gene mRNA expression profile in circulating tumor cells predicts response to aromatase inhibitors in metastatic breast cancer patients</p>	<p>Minimally-invasive Expression in Circulating Tumor Cells from Metastatic Breast Cancer Patients Enrolled in North Central Cancer Treatment Group Trials. N00917322/1521197</p>
<p>2010 2014 2012</p>	<p>2005</p>	<p>2010</p>	<p>2012</p>	<p>2015</p>	<p>2014</p>	<p>2016</p>	<p>2011</p>
<p>54 / 53 54 34</p>	<p>60</p>	<p>76</p>	<p>98</p>	<p>39</p>	<p>66</p>	<p>78</p>	<p>146</p>
<p>Case-Control Case series Cell line</p>	<p>Case series</p>	<p>Case series</p>	<p>Case series</p>	<p>Case series</p>	<p>Case series</p>	<p>Case series</p>	<p>Case series</p>
<p>Stage I-III breast cancer Early breast cancer patients breast cancer</p>	<p>BC</p>	<p>Advanced BC</p>	<p>metastatic breast cancer</p>	<p>MBC</p>	<p>CTCs positive breast cancer</p>	<p>MBC</p>	<p>MBC</p>
<p>Detection of invasive CTCs and Detection of CTCs, Molecular phenotyping of circulating Enumeration and characterization of CTCs CTCs enumeration and characterization</p>	<p>Enumeration and characterization of CTCs</p>	<p>Enumeration and characterization of CTCs</p>	<p>Enumeration and characterization of CTCs</p>	<p>CTCs enumeration and molecular characterization (Whole Blood PIK3CA)</p>	<p>comprehensive molecular characterization of CTCs</p>	<p>CTC gene expression profile identification (RNA isolation from CTCs, qRT-PCR and quantification of gene transcripts) and Enumeration</p>	<p>Enumeration and molecular characterization of CTCs</p>
<p>Collagen adhesion matrix (CAM) assay, CAM uptake assay, PCR Adm Test/BreastCancer™ (AdmGen AG, Germany), qPCR gene expression analysis qRT-PCR, Tissue microarrays (TMAs) at CellSearch Epithelial Cell Kit (Veridex LLC), stamboatkin-1 (STC-1), N-acetylglucosaminyltransferase (GalNAcT), and metastoma antigen gene family-A3 (MAGE-A3) EPCAM, HER2, MUC1, TOP1, TOP2A, CTSD, ST6, CK19 (EPCAM) and selection (CK8/18/19) markers used in this method. While CD146 can detect EPCAM-negative CTCs, we have evaluated the value of various cytokeratins and CD19f to detect CK8/18/19-negative CTCs Adjuvant chemotherapy, Neo-adjuvant chemotherapy Neoadjuvant and/or adjuvant chemotherapy</p>	<p>Immune system (Cytokeratin, Epithelial Cell Kit, FerriGold, Frickhausen, Germany), standard ELISA, DMK, CA15.3 second generation by Abbott CA15.3, epithelial cell adhesion molecule (EPCAM), Ki-67 antigen, HER-2/neu expression, estrogen receptor, progesterone receptor, androgen receptor, and Herceptin. Anti HER-2/neu antibody trastuzumab Chemotherapy</p>	<p>immunomagnetic separation using FerriGold nanoparticles binding anti-epithelial cell adhesion molecules (EPCAM) and EPCAM+, CK+, DAPI+, CD45+, and HER2/neu+.</p>	<p>fluorescence in situ hybridization FISH, immunomagnetic techniques using magnetic beads, immunocytochemical methods Estrogen receptor, Progesterone receptor Epidermal growth factor receptor (EGFR), HER2 and TOP2A</p>	<p>Combination of the CellSearch and DEParity technologies Whole Genome Amplification (WGA) and sequencing analysis Heterogeneity of PIK3CA mutational status within single CTCs</p>	<p>DEParityTM technology (Sifon Bioscience, SpA), Whole genome amplification (WGA) using the AmpliTm WGA Kit</p>	<p>The CellTracks Analyzer (Veridex LLC), quantitative reverse transcriptase polymerase chain reaction (q-PCR) 8 genes: TWIST1, KRTR1, PTFR, EEF1A2, PTPRK, EGFR, CXCL4, HERB3</p>	<p>Reverse transcription/quantitative PCR by a BioRad/CyberIQ Lapatinib +Cetuximab, Sorafenib, Imatinib +cetuximab and poliguanine + capcitabine CK19, MGB1, and b2-microglobulin (B2M) mRNA levels</p>
<p>CTC detection may be a promising early marker of disease progression potentially enhancing the clinical therapeutic decisions The novel cytokeratins provided no substantial benefit, but adding CD19f to CK8/18/19 as a selection marker resulted in improved recovery of normal-like epithelial cells Combined staining of CK8/18/19 and CD19f after CD146/EPCAM enrichment is likely to further improve</p>	<p>Patients with HER2 overexpression in CTCs had poorer survival compared with those without CTCs or with HER2-CTCs</p>	<p>monitoring since heterogeneity of the biomarker distribution in CTCs and the lack of correlation with the primary tumor biomarker status were found. Further</p>	<p>targeted therapy (namely, trastuzumab)</p>	<p>Target therapy</p>	<p>amplification) and (10) genome-wide analysis using hybrid CTCs uncovered pre-existing cells resistant</p>	<p>Chemotherapy, combined with a type of targeted therapy such as Trastuzumab, Tamoxifen therapy, discriminates good and poor outcome to first-line aromatase inhibitors in MBC patients. Although results need to be validated, this study underscores the</p>	<p>response to therapy of</p>

Continuance of Table 2.

Reichardt, S.(21) Soliani S,(68) Spilloni, M(69), Tewes, M.(133)	Patients with either large operable or locally advanced tumors, tumors with negative hormone receptor status, or receptor-positive tumors but clinically node-positive disease stage I to III early breast cancer MBC	Case series Case/control Case series Case series	2010 2015 2014 2009	287 21,730 122 42,788	468	221	42	82,51	Waltherer, Markus(134)	The prognostic impact of circulating tumor cells in subtypes of metastatic breast cancer
Wang, H. Y.(135)	Patients with either large operable or locally advanced tumors, tumors with negative hormone receptor status, or receptor-positive tumors but clinically node-positive disease stage I to III early breast cancer MBC	Case series in	2013	2014	2006	2006	2006	2006	Walling, P.(102)	HER2-positive circulating tumor cells indicate poor clinical outcome in stage I to III breast cancer patients
Zakheim, B. K.(136)	Patients with either large operable or locally advanced tumors, tumors with negative hormone receptor status, or receptor-positive tumors but clinically node-positive disease stage I to III early breast cancer MBC	Case/control	2006	2006	2006	2006	2006	2006	Zakheim, B. K.(136)	Detection of circulating tumor cells in peripheral blood of breast cancer patients during or after therapy using a multiplex real-time RT-PCR assay
Food and Drug Administration-approved CellSearch system for CTC detection and evaluation of HER2 expression and developed HER2 immunoscreening for CTC SYBR green-based real-time quantitative polymerase chain reaction assays cell lines SKBR3 and MDAMB231 were obtained from the American Type Culture Collection (Manassas, VA, USA), Fcolli-Hyqque density gradient (Madison, VA, USA), Fcolli-Hyqque density gradient (Madison, VA, USA) and progesterone (PR) receptor expression was assessed by RT-PCR	CTCs characterization CTCs detection and characterization, CTCs Cell culture Molecular characterization of single CTCs	CellSearch system (Veridex, LLC, Warren, NJ, USA), Immunohistochemical staining (IHC)	Cell culture quantitative reverse transcription PCR (RT-qPCR) and RT-qPCR/EpCAM assay	EpCAM, cytokeratin (CK) 19, human epidermal growth factor (HER) 2, K67, human telomerase reverse transcriptase (hTERT) and vimentin	Estrogen receptor and Her2	postoperative ipsilateral adjuvant therapy, cytarosine	Chemotherapy + hormone therapy, Herceptin, No treatment and	expression levels of tamoxifen, B305D, gamma-aminobutyrate type A receptor pi subunit (GABA _A π), GABRB2 and B726p	Wang, H. Y.(135)	Detection of circulating tumor cells in patients with breast cancer using the quantitative RT-PCR assay for monitoring of therapy efficacy
Food and Drug Administration-approved CellSearch system for CTC detection and evaluation of HER2 expression and developed HER2 immunoscreening for CTC SYBR green-based real-time quantitative polymerase chain reaction assays cell lines SKBR3 and MDAMB231 were obtained from the American Type Culture Collection (Manassas, VA, USA), Fcolli-Hyqque density gradient (Madison, VA, USA), Fcolli-Hyqque density gradient (Madison, VA, USA) and progesterone (PR) receptor expression was assessed by RT-PCR	CTCs characterization CTCs detection and characterization, CTCs Cell culture Molecular characterization of single CTCs	CellSearch system (Veridex, LLC, Warren, NJ, USA), Immunohistochemical staining (IHC)	Cell culture quantitative reverse transcription PCR (RT-qPCR) and RT-qPCR/EpCAM assay	EpCAM, cytokeratin (CK) 19, human epidermal growth factor (HER) 2, K67, human telomerase reverse transcriptase (hTERT) and vimentin	Estrogen receptor and Her2	postoperative ipsilateral adjuvant therapy, cytarosine	Chemotherapy + hormone therapy, Herceptin, No treatment and	expression levels of tamoxifen, B305D, gamma-aminobutyrate type A receptor pi subunit (GABA _A π), GABRB2 and B726p	Walling, P.(102)	HER2-positive circulating tumor cells indicate poor clinical outcome in stage I to III breast cancer patients
Food and Drug Administration-approved CellSearch system for CTC detection and evaluation of HER2 expression and developed HER2 immunoscreening for CTC SYBR green-based real-time quantitative polymerase chain reaction assays cell lines SKBR3 and MDAMB231 were obtained from the American Type Culture Collection (Manassas, VA, USA), Fcolli-Hyqque density gradient (Madison, VA, USA), Fcolli-Hyqque density gradient (Madison, VA, USA) and progesterone (PR) receptor expression was assessed by RT-PCR	CTCs characterization CTCs detection and characterization, CTCs Cell culture Molecular characterization of single CTCs	CellSearch system (Veridex, LLC, Warren, NJ, USA), Immunohistochemical staining (IHC)	Cell culture quantitative reverse transcription PCR (RT-qPCR) and RT-qPCR/EpCAM assay	EpCAM, cytokeratin (CK) 19, human epidermal growth factor (HER) 2, K67, human telomerase reverse transcriptase (hTERT) and vimentin	Estrogen receptor and Her2	postoperative ipsilateral adjuvant therapy, cytarosine	Chemotherapy + hormone therapy, Herceptin, No treatment and	expression levels of tamoxifen, B305D, gamma-aminobutyrate type A receptor pi subunit (GABA _A π), GABRB2 and B726p	Zakheim, B. K.(136)	Detection of circulating tumor cells in peripheral blood of breast cancer patients during or after therapy using a multiplex real-time RT-PCR assay
Epirubicin/cyclophosphamide prior to randomization to docetaxel alone, docetaxel in combination with capecitabine, docetaxel followed by capecitabine and additional trastuzumab treatment Adjuvant chemotherapy Chemotherapy An anthracycline- or taxane-based chemotherapy, trastuzumab	HER2 expression on CTC. CK-19 gene expression Apoptotic markers: K67 and M30 status of CTCs HER2, MUC1 and GAT3-2 transcript	HER2 targeting therapy, HER2 directed treatment combined with chemotherapy	Adjuvant chemotherapy, neoadjuvant chemotherapy	detection of CTC-related markers. Data from this study suggest that RT-qPCR assay of CTC markers might be useful in selecting appropriate	HER2-targeting therapy, HER2 directed treatment combined with chemotherapy	Adjuvant chemotherapy, neoadjuvant chemotherapy	Chemotherapy + hormone therapy, Herceptin, No treatment and	expression levels of tamoxifen, B305D, gamma-aminobutyrate type A receptor pi subunit (GABA _A π), GABRB2 and B726p	Walling, P.(102)	HER2-positive circulating tumor cells indicate poor clinical outcome in stage I to III breast cancer patients
A study for determining CTC biomarkers in human breast cancer women that suggest that the CK-19 mRNA expression investigation may be useful for monitoring CTCs in the blood of metastatic breast cancer patients predicting early metastatic relapse or monitoring of anti-metastatic treatments. The apoptotic index of CTCs is increased during clinical dormancy, whereas the proliferation index is increased on relapse. In addition, apoptotic CTCs are more frequently encountered during follow-up in DC patients who remain disease-free compared to those with subsequent late relapse, suggesting that monitoring proliferation and apoptosis in CTCs during clinical dormancy merits further investigation as a tool for predicting late disease recurrence.	HER2 expression on CTC. CK-19 gene expression Apoptotic markers: K67 and M30 status of CTCs HER2, MUC1 and GAT3-2 transcript	HER2 targeting therapy, HER2 directed treatment combined with chemotherapy	Adjuvant chemotherapy, neoadjuvant chemotherapy	detection of CTC-related markers. Data from this study suggest that RT-qPCR assay of CTC markers might be useful in selecting appropriate	HER2-targeting therapy, HER2 directed treatment combined with chemotherapy	Adjuvant chemotherapy, neoadjuvant chemotherapy	Chemotherapy + hormone therapy, Herceptin, No treatment and	expression levels of tamoxifen, B305D, gamma-aminobutyrate type A receptor pi subunit (GABA _A π), GABRB2 and B726p	Zakheim, B. K.(136)	Detection of circulating tumor cells in peripheral blood of breast cancer patients during or after therapy using a multiplex real-time RT-PCR assay

They indicated that checking of Her2 expression on CTCs might be beneficial in trials with anti-Her2 (48) or optimizing individually tailored therapies in Her2-positive MBC patients (49), also the finding of Her2 mRNA-positive CTCs after the adjuvant chemotherapy completion possibly will provide clinically useful data concerning the efficacy of treatment and operable breast cancer prognosis (50). Her2-positive CTCs count will be required to compare the assay-dependent Her2 status of CTCs to the clinical response to Her2-targeted therapies (48,51,52) because patients with Her2 overexpression in CTCs taken inferior progression-free survival compared with those without CTCs or with Her2-CTCs (53).

Lapatinib, which may be given with the chemotherapy drug capecitabine (Xeloda) or a biological therapy called trastuzumab (Herceptin) is an effective drug in decreasing Her2-positive CTCs in patients with MBC irrespectively of the Her2 status of the primary tumor (54) but in one reported case it is shown that expression of epidermal growth factor receptor (EGFR) could predict response to lapatinib-based treatments (28). The association between EGFR-positive CTCs and Luminal tumors was justified in one study (47).

In one research whole genome amplification 3-5 single CTCs per patient were analyzed by next generation sequencing (NGS) for fifty cancer-related genes (55).

They found 51 sequence variants in 25 genes including both inter- and intra-patient heterogeneity in the mutational status of CTCs. The highest number of somatic deleterious mutations was found in the gene TP53, whose mutation is associated by means of adverse prognosis in breast cancer and supports the applicability of a non-invasive approach based on the liquid biopsy in MBC patients for the development of new therapeutic strategies in precision medicine. Checking the status of eight genes mRNA expression profile in circulating tumor cells conducted by Reijm E.A., *et al.*, identified that 75% most variable genes which are differentially expressed in two groups of good and poor responders and was significantly associated with outcome (56). This predictor recognized poor responding patients with a sensitivity of 63% and a positive predictive value of 75%, whereas good responding patients were properly predicted in 85% of the cases.

Some of other studied molecule markers are

- Carcino Embryonic Antigen (CEA) and Cancer Antigen 15-3 (CA15-3) amount combination can predict survival (OS and PFS) (33,57-60). Independent prognostic significance of elevated preoperative serum CEA and CA15-3 levels were reconfirmed in Luminal B breast cancer (60,61).
- Interleukin-4,-5,-6,-8-13, Th2 cytokines (62) In CTC-negative patients, expression of interleukin-8 (IL-8) and IL-13 had increased on the occasion of being negative for progesterone receptor. IL-5 was significantly enlarged in patients with human epidermal growth factor receptor 2 (Her2)-positive and lymph node-positive, IL-4 was increased in patients with progesterone receptor-positive and estrogen receptor-negative, in addition, IL-6 levels was escalated in patients with tumor grade G3 lacking progesterone receptor expression. Th2 cytokines are expressively changed in patients who were CTC-negative and progesterone receptor-positive consequently IL-4 plays a leading role in the poor outcome of a number of breast cancer cases (62).
- Stanniocalcin-1 (STC-1), N-acetyl galactosaminyl transferase (GalNacT), and melanoma antigen gene family-A3 (MAGE-A3) assessment by quantitative Real Time (qRT) PCR for mRNA expression showed a correlation between the total axillary LN (ALN), non-sentinel lymph node (SLN) and SLN histopathology status. So the recognition of CTCs proposes an innovative means to assess the presence of systemic disease spreading relative to SLN and ALN histopathology status (63,64).
- The immune checkpoint regulators such as PD-L1 (CD279), PD-L2 (B7-DC; CD273), reported the expression of PD-L1 on CTCs and CTC/PD-L1 assay as a useful screening for liquid biopsy in future clinical trials for stratification and monitoring of cancer patients undergoing immune checkpoint blockade (53,65).
- MUC1, TOP1, TOP2A, CTSD, ST6, CK19 as a promising early marker of disease progression which is useful for both on behalf of both the prediction of outcome and checking the effect of treatment (66).
- Cytokeratin 7, 8, 18 and 19 (67) to predicting early metastatic relapse or monitoring of anti-metastasis treatments (68).
- Apoptotic markers like Ki67 and M30 (69) that are enlarged during clinical dormancy, but the proliferation index is augmented on relapse or late disease recurrence (69).
- Hotspot mutations in ESR1, phosphatidylinositol-4, 5-bisphosphate 3-kinase, catalytic subunit alpha (PIK3CA), tumor protein p53 (TP53), fibroblast growth factor receptor 1 (FGFR1), and fibroblast growth factor receptor 2 (FGFR2) for termination of ineffective endocrine therapies and substituting another treatment (70).
- Circulating levels transforming growth factor-beta (TGF-β) and Chemokine (C-X-C Motif) Ligand-1 (CXCL1) which are linked to the poor prognosis, besides lung metastases in patients with breast cancer (71).

Discussion

There has been a growing interest in exploring the clinical significance of CTCs in personalized diagnosis and treatment of breast cancer over the last decade. Here, we report the first systematic review of published studies evaluating the association of CTCs enumeration and molecular identification with clinicopathological characteristics and clinical outcome breast cancer. We identified sixty-nine studies. One of them was related to the best techniques for detection the early markers of response to chemotherapy which may ultimately lead to tailored therapies and avoid cumulative toxicity, using metabolic imaging with [18F] 3'-deoxy-3'-fluorothymidine PET (FLT-PET) in women with advanced breast cancer, before and during docetaxel therapy could provide a powerful, albeit expensive, tool to assess immediate responses to therapy (37). Another study was related to using of whole-body FDG-PET/CT in MBC patients who has relapsed/progressive MBC. It is

Circulating tumor cells

shown that existence of widespread bone metastases identified by FDG-PET/CT is connected directly to the increased CTC numbers in MBC (30). Recently some methods have been established for constantly identifying and quantifying CTCs in blood samples (72-75). Breakthrough in the biosensor field and microfluidic chip for discriminating separation of circulating tumor cells (CTCs) recently have brought the new insight for tracking metastatic breast cancer, CTCs enrichment and isolation platforms (76-79).

Sixty-seven studies were mainly focused on CTC isolation, enumeration and characterization before and during therapy to estimate the utility in changing therapy against maintaining therapy in breast cancer patients. Most assays established for the enumeration of CTCs by means of CellSearch system which rely on the expression of the epithelial cell adhesion molecule (EpCAM). EpCAM is a transmembrane glycoprotein mediating Ca^{2+} -independent cell adhesion molecule in epithelial which also is involved in cell signaling, migration, proliferation, and differentiation (80-82). The weak point of this method is that may not detect CTCs that express no/low levels of EpCAM like cells which are undergoing epithelial-to-mesenchymal transition (EMT) (83), therefore estimated the value of several cytokeratin and CD49f to distinguish CK8/18/19- negative CTCs. For further improvement of CTC detection in breast cancer shared staining of CK8/18/19 and CD49f following CD146/EpCAM enrichment is suggested (84). Furthermore, a functional cell separation method, called collagen adhesion matrix (CAM) assay, has described recently to improve enrichment and identification steps methods (85-87).

Although CTC status was prognostic and changing CTC levels during chemotherapy are useful to monitor therapy efficacy (20,30,31,34,39-41,43,58,88-93), simple enumeration has a low predictive value and cannot predict a specific course of treatment (94,95), and it needs to be added to full clinic-pathological predictive models (88). The early DETECT trials revealed that a serial CTC measurements before and after chemotherapy shown a prognostic value (42,57) but subsequent related trials evaluating targeted agents based on phenotypes of CTCs (96). Molecular characterization of CTCs is an important step forward to the way of personalizing management of breast cancer to inform the discovery of exact therapeutic predictors. Because of high circulating tumor cell (CTC) heterogeneity (97), it can easily say that there is an extreme need for molecular profiling of CTCs including protein expression, phenotypic changes and gene expression (48,98). It has been shown that treatment

efficiency or recurrent of breast tumors (MBC or TNBC) could be predictable with analysis the expression of some molecules like a proto-oncogene Her2 (21,39,50,54,89,99). The result of these studies indicates that after the accomplishment of adjuvant chemotherapy, detection of Her2 positive CTCs may provide clinically useful information related to the treatment efficacy (50). The clinical trial Gepar Quattro combined neoadjuvant (NT) attitudes (epirubicin/cyclophosphamide prior to randomization to docetaxel alone, docetaxel in combination with capecitabine, or docetaxel followed by capecitabine) plus additional trastuzumab treatment in patients who have Her2-positive tumors then shown that CTCs detection had not been connected to the primary tumor characteristics, but CTC Her2 overexpression was limited to ductal carcinomas and was completely connected to the higher tumor stage (21). CTC numbers were truncated in patients with primary breast cancer, in addition, the reduction in CTCs amount during treatment was not related to the standard clinical characteristics and primary tumor response so the evidence of the CTCs Her2 might be beneficial for Her2-directed therapies monitoring (21). Moreover several studies checked the prognostic impact of Her2 in combination with some other cellular markers like hormone receptors (ER and PR) and Her2 expression (45,46,55,100-102), epidermal growth factor receptor (EGFR) and Her2 in reaction to a treatment regime comprising lapatinib (a dual EGFR and Her2 tyrosine kinase inhibitor) (28). EGFR-positive CTCs were associated with Luminal tumors in a patient who is impressed by chemorefractory metastatic Her2-positive breast cancer receiving lapatinib (47). Disease progress was completely connected to a recurrence in CTCs; representing EGFR expression could calculate a response to lapatinib-based treatments (28).

CTCs prognostic outcome was fewer evident in Her2 positive MBC patients cured by targeted therapy (45), which support this idea that the quantity of CTCs, together with the biologic characteristics, desires to be wisely taken into account in the future analysis. In non-metastatic breast cancer CTC biomarker analysis more than Her2 might be useful as a replacement marker for therapeutic selection and monitoring since heterogeneity of the biomarker distribution in CTCs and the lack of correlation with the primary tumor biomarker status were found (47). By way of illustration a trial which checked the multidrug-resistance-related proteins (MRPs), aldehyde dehydrogenase 1 (ALDH1), estrogen receptor an (ERa) plus Her2/neu, indicated to the existence of CTCs expressing MRPs and ALDH1, is prognostic for chemotherapy response in MBC patients (43). A

difference in PFS was obvious in two groups of CTCs+and CTCs-patients that were undersized in patients with a drug resistance CTCs profile and in patients who has expressed two or more MRPs on their CTCs, so the existence of CTCs expressing MRPs and ALDH1 stands prognostic for chemotherapy (22,103).

Four markers (EPCAM, CD47, CD44 and MET) which are known to be involved in tumor genesis (104, 105) and are co-regulated with the TGF- β signaling pathway (106) has been checked in the earliest stage of breast cancer to plan intervention settings that modify the patient-specific survival prospect (29). Through a branching process model, the survival times and this four markers gene expression correlation can predict personalized OS or PFS especially drugs such as bisphosphonates. The analysis of circulating tumor cells effects on the disease progression offering a quantitative measurement of the cell driver mutations which are responsible for invading the bone tissue. This model lets to plan intervention scenarios that adjust the patient-specific survival chance by altering the populations of circulating tumor cells, in addition, the situation could be extended to other cancer metastasis dynamics (107).

Thanks to several advancements in molecular genetics technology like high-throughput NGS, multi-gene mutation analysis that provides comprehensive genetic information on breast cancer molecular pathology, make it much easier to find a precision and more effective therapeutic targets (108). Two studies, evaluating genomic alterations in cancer-related genes of CTCs to provide insights into mechanisms of tumor metastases and drug resistance (55,56). It has been shown that CTC characteristics are more closely linked to the dynamic modifications of the disease status and CTCs genetic analysis is a non-invasive approach based on the liquid biopsy in metastatic breast cancer patients which, in perspective, should allow investigating the clonal evolution of the tumor for the development of new therapeutic strategies in precision medicine (55). Some researchers indicated to the fact that NGS in combination with Fluorescence-activated cell sorting (FACS) and Immunohistochemistry (IHC) is an excellent way to outline copy number in a single cell in several cancer types, as well as breast cancer (46,109-111). Also, single cell analysis has identified the clinically significant genomic difference between primary tumors and CTCs (112-114) offer fundamental information for personalized treatment decisions and shed light on drug resistance and tumor heterogeneity mechanisms (114).

Especially for individualized testing of the drug some studies have been working on in vitro and in vivo CTCs

culture (115,116) and assessed genemutations in circulating tumor cell from cancer patients by next generation sequencing (NGS) (55,117). Mutation detection in PIK3CA, FGFR2, and ESR-1 through CTC-iChip in breast cancer patients and drug sensitivity testing revealed that the selective estrogen receptor modulators (SERMs) tamoxifen and raloxifene, and the selective ER degrader (SERD) fulvestrant, were ineffective in ESR-1 mutant cells. Cultured CTCs were highly sensitive to the PIK3CA inhibitor BYL719 and the FGFR2 inhibitor AZD4547 (118). The enumeration of CTCs using Cell Search system and CAM assay which rely on the expression of the cell surface marker (EpCAM, CD49f, CD146/EpCAM enrichment) during or after treatment is mostly beneficial for predicting early metastatic relapse.

In conclusion it can be said that the clinical significance of CTCs molecular profiling and characteristics is more accurate than CTCs enumeration before and during treatment, especially for making the best personalized treatment decision CTCs molecular markers like Her2, EGFR, CEA, CA15-3, CK19, Ki67, PIK3CA, TGF- β , and CXCL1 are really valuable to be checked.

References

1. National Comprehensive Cancer Network Clinical Practice Guidelines in Oncology. (Accessed December 2016, 30, at http://www.nccn.org/professionals/physician_gls/PDF/breast.pdf).
2. ACS Research Updates, 10 Must-Know 2015 Global Cancer Facts. American Cancer Society. 2015. (Accessed December 2016, 30, at <https://www.cancer.org/latest-news/10-must-know-2015-global-cancer-facts.html>).
3. Perou CM, Sorlie T, Eisen MB, van de Rijn M, Jeffrey SS, Rees CA, et al. Molecular portraits of human breast tumours. *Nature* 2000;406:747-52.
4. Sorlie T, Perou CM, Tibshirani R, Aas T, Geisler S, Johnsen H, et al. Gene expression patterns of breast carcinomas distinguish tumor subclasses with clinical implications. *Proc Natl Acad Sci U S A* 2001;98:10869-74.
5. Gort M, Broekhuis M, Otter R, Klazinga NS. Improvement of best practice in early breast cancer: actionable surgeon and hospital factors. *Breast Cancer Res Treat* 2007;102:219-26.
6. Elston CW, Pinder SE. Prognostic factors in invasive carcinoma of the breast. *Clin Oncol (R Coll Radiol)* 1998;10:3.
7. Curtis C, Shah SP, Chin S-F, Turashvili G, Rueda OM, Dunning MJ, et al. The genomic and transcriptomic

Circulating tumor cells

- architecture of 2,000 breast tumours reveals novel subgroups. *Nature* 2012;486:346-52.
8. Milioli HH, Vimieiro R, Riveros C, Tishchenko I, Berretta R, Moscato P. The Discovery of Novel Biomarkers Improves Breast Cancer Intrinsic Subtype Prediction and Reconciles the Labels in the METABRIC Data Set. *PLoS ONE* 2015;10:e0129711.
 9. Van Poznak C, Somerfield MR, Bast RC, Cristofanilli M, Goetz MP, Gonzalez-Angulo AM, et al. Use of biomarkers to guide decisions on systemic therapy for women with metastatic breast cancer: American Society of Clinical Oncology Clinical Practice Guideline. *J Clin Oncol* 2015;20:2695-704.
 10. Ross JS, Symmans WF, Pusztai L, Hortobagyi GN. Breast cancer biomarkers. *Adv Clin Chem* 2004;40:99-125.
 11. Martelotto LG, Ng CK, Piscuoglio S, Weigelt B, Reis-Filho JS. Breast cancer intra-tumor heterogeneity. *Breast Cancer Res* 2014;16:1-11.
 12. Gerlinger M, Rowan AJ, Horswell S, Larkin J, Endesfelder D, Gronroos E, et al. Intratumor heterogeneity and branched evolution revealed by multiregion sequencing. *N Engl J Med* 2012;366:883-92.
 13. Bedard PL, Hansen AR, Ratain MJ, Siu LL. Tumour heterogeneity in the clinic. *Nature* 2013;501:355-64.
 14. Pantel K, Alix-Panabières C. Real-time liquid biopsy in cancer patients: fact or fiction? *Cancer Res* 2013;73:6384-8.
 15. Sun YF, Yang XR, Zhou J, Qiu SJ, Fan J, Xu Y. Circulating tumor cells: advances in detection methods, biological issues, and clinical relevance. *J Cancer Res Clin Oncol* 2011;137:1151-73.
 16. Allard WJ, Matera J, Miller MC, Repollet M, Connelly MC, Rao C, et al. Tumor cells circulate in the peripheral blood of all major carcinomas but not in healthy subjects or patients with nonmalignant diseases. *Clin Cancer Res* 2004;10:6897-904.
 17. Cristofanilli M, Budd GT, Ellis MJ, Stopeck A, Matera J, Miller MC, et al. Circulating tumor cells, disease progression, and survival in metastatic breast cancer. *N Engl J Med* 2004;351:781-91.
 18. Bidard FC, Mathiot C, Delaloue S, Brain E, Giachetti S, de Cremoux P, et al. Single circulating tumor cell detection and overall survival in nonmetastatic breast cancer. *Ann Oncol* 2010;21:729-33.
 19. Xenidis N, Ignatiadis M, Apostolaki S, Perraki M, Kalbakis K, Agelaki S, et al. Cytokeratin-19 mRNA-positive circulating tumor cells after adjuvant chemotherapy in patients with early breast cancer. *J Clin Oncol* 2009;27:2177-84.
 20. Cristofanilli M, Hayes DF, Budd GT, Ellis MJ, Stopeck A, Reuben JM, et al. Circulating tumor cells: a novel prognostic factor for newly diagnosed metastatic breast cancer. *J Clin Oncol* 2005;23:1420-30.
 21. Riethdorf S, Muller V, Zhang L, Rau T, Loibl S, Komor M, et al. Detection and HER2 expression of circulating tumor cells: prospective monitoring in breast cancer patients treated in the neoadjuvant GeparQuattro trial. *Clin Cancer Res* 2010;16:2634-45.
 22. Gradilone A, Naso G, Raimondi C, Cortesi E, Gandini O, Vincenzi B, et al. Circulating tumor cells (CTCs) in metastatic breast cancer (MBC): prognosis, drug resistance and phenotypic characterization. *Ann Oncol* 2011;22:86-92.
 23. Hayes DF, Smerage J. Is there a role for circulating tumor cells in the management of breast cancer? *Clin Cancer Res* 2008;14:3646-50.
 24. Giuliano M, Herrera S, Christiny P, Shaw C, Creighton CJ, Mitchell T, et al. Circulating and disseminated tumor cells from breast cancer patient-derived xenograft-bearing mice as a novel model to study metastasis. *Breast Cancer Res* 2015;17:3.
 25. Toss A, Mu Z, Fernandez S, Cristofanilli M. CTC enumeration and characterization: moving toward personalized medicine. *Ann Trans Med* 2014;2:108.
 26. Smith BM, Slade MJ, English J, Graham H, Luchtenborg M, Sinnott HD, et al. Response of circulating tumor cells to systemic therapy in patients with metastatic breast cancer: comparison of quantitative polymerase chain reaction and immunocytochemical techniques. *J Clin Oncol* 2000;18:1432-9.
 27. Liu MC, Shields PG, Warren RD, Cohen P, Wilkinson M, Ottaviano YL, et al. Circulating tumor cells: a useful predictor of treatment efficacy in metastatic breast cancer. *J Clin Oncol* 2009;27:5153-9.
 28. Liu Z, Fusi A, Schmittel A, Tinhofer I, Schneider A, Keilholz U. Eradication of EGFR-positive circulating tumor cells and objective tumor response with lapatinib and capecitabine. *Cancer Biol Ther* 2010;10:860-4.
 29. Ascolani G, Occhipinti A, Lio P. Modelling circulating tumour cells for personalised survival prediction in metastatic breast cancer. *PLoS Comput Biol* 2015;11:e1004199.
 30. De Giorgi U, Valero V, Rohren E, Mego M, Doyle GV, Miller MC, et al. Circulating tumor cells and bone metastases as detected by FDG-PET/CT in patients with metastatic breast cancer. *Ann Oncol* 2010;21:33-9.
 31. Green TL, Cruse JM, Lewis RE, Craft BS. Circulating tumor cells (CTCs) from metastatic breast cancer patients linked to decreased immune function and response to treatment. *Exp Mol Pathol* 2013;95:174-9.
 32. Cristofanilli M, Budd GT, Ellis MJ, Stopeck A, Matera J,

- Miller MC, et al. Circulating tumor cells, disease progression, and survival in metastatic breast cancer. *N Engl J Med* 2004;351:781-91.
33. Bidard FC, Peeters DJ, Fehm T, Nole F, Gisbert-Criado R, Mavroudis D, et al. Clinical validity of circulating tumour cells in patients with metastatic breast cancer: a pooled analysis of individual patient data. *Lancet Oncol* 2014;15:406-14.
 34. Bidard F-C, Mathiot C, Delalogue S, Brain E, Giachetti S, de Cremoux P, et al. Single circulating tumor cell detection and overall survival in nonmetastatic breast cancer. *Ann Oncol* 2010;21:729-33.
 35. Pierga JY, Hajage D, Bachelot T, Delalogue S, Brain E, Campone M, et al. High independent prognostic and predictive value of circulating tumor cells compared with serum tumor markers in a large prospective trial in first-line chemotherapy for metastatic breast cancer patients. *Ann Oncol* 2012;23:618-24.
 36. Pierga JY, Bidard FC, Mathiot C, Brain E, Delalogue S, Giachetti S, et al. Circulating tumor cell detection predicts early metastatic relapse after neoadjuvant chemotherapy in large operable and locally advanced breast cancer in a phase II randomized trial. *Clin Cancer Res* 2008;14:618-24.
 37. Contractor K, Aboagye EO, Jacob J, Challapalli A, Coombes RC, Stebbing J. Monitoring early response to taxane therapy in advanced breast cancer with circulating tumor cells and [18F] 3'-deoxy-3'-fluorothymidine PET: a pilot study. *Biomark Med* 2012;6:231-3.
 38. De Giorgi U, Valero V, Rohren E, Dawood S, Ueno NT, Miller MC, et al. Circulating tumor cells and [18F]fluorodeoxyglucose positron emission tomography/computed tomography for outcome prediction in metastatic breast cancer. *J Clin Oncol* 2009;27:3303-11 .
 39. Hall C, Karhade M, Laubacher B, Anderson A, Kuerer H, DeSynder S, et al. Circulating Tumor Cells After Neoadjuvant Chemotherapy in Stage I-III Triple-Negative Breast Cancer. *Ann Surg Oncol* 2015;22:S552-8 .
 40. Karhade M, Hall C, Mishra P, Anderson A, Kuerer H, Bedrosian I, et al. Circulating tumor cells in non-metastatic triple-negative breast cancer. *Breast Cancer Res Treat* 2014;147:325-33 .
 41. Hall CS, Karhade M, Laubacher BA, Kuerer HM, Krishnamurthy S, DeSnyder S, et al. Circulating Tumor Cells and Recurrence After Primary Systemic Therapy in Stage III Inflammatory Breast Cancer. *J Natl Cancer Inst* 2015;107:djv250.
 42. Müller V, Riethdorf S, Rack B, Janni W, Fasching PA, Solomayer E, et al. Prognostic impact of circulating tumor cells assessed with the CellSearch System™ and AdnaTest Breast™ in metastatic breast cancer patients: the DETECT study. *Breast Cancer Res* 2012;14:1-8.
 43. Giuliano M, Giordano A, Jackson S, Hess KR, De Giorgi U, Mego M, et al. Circulating tumor cells as prognostic and predictive markers in metastatic breast cancer patients receiving first-line systemic treatment. *Breast Cancer Res* 2011;13:1-9.
 44. Giordano A, Giuliano M, De Laurentiis M, Arpino G, Jackson S, Handy BC, et al. Circulating tumor cells in immunohistochemical subtypes of metastatic breast cancer: lack of prediction in HER2-positive disease treated with targeted therapy. *Ann Oncol* 2012;23:1144-50.
 45. Giordano A, Giuliano M, De Laurentiis M, Eleuteri A, Iorio F, Tagliaferri R, et al. Artificial neural network analysis of circulating tumor cells in metastatic breast cancer patients. *Breast Cancer Res Treat* 2011;129:451-8.
 46. Peeters DJ, van Dam PJ, Van den Eynden GG, Rutten A, Wuyts H, Pouillon L, et al. Detection and prognostic significance of circulating tumour cells in patients with metastatic breast cancer according to immunohistochemical subtypes. *Br J Cancer* 2014;110:375-83 .
 47. Nadal R, Fernandez A, Sanchez-Rovira P, Salido M, Rodriguez M, Garcia-Puche JL, et al. Biomarkers characterization of circulating tumour cells in breast cancer patients. *Breast Cancer Res* 2012;14:R71.
 48. Ignatiadis M, Rothe F, Chaboteaux C, Durbecq V, Rouas G, Criscitiello C, et al. HER2-positive circulating tumor cells in breast cancer. *PLoS One* 2011;6:e15624 .
 49. Ligthart ST, Coumans FAW, Attard G, Mulick Cassidy A, de Bono JS, Terstappen LWMM. Unbiased and automated identification of a circulating tumour cell definition that associates with overall survival. *Plos One* 2011;6:e27419.
 50. Apostolaki S, Perraki M, Pallis A, Bozionelou V, Agelaki S, Kanellou P, et al. Circulating HER2 mRNA-positive cells in the peripheral blood of patients with stage I and II breast cancer after the administration of adjuvant chemotherapy: evaluation of their clinical relevance. *Ann Oncol* 2007;18(5):851-8.
 51. Fehm T, Becker S, Duerr-Stoerzer S, Sotlar K, Mueller V, Wallwiener D, et al. Determination of HER2 status using both serum HER2 levels and circulating tumor cells in patients with recurrent breast cancer whose primary tumor was HER2 negative or of unknown HER2 status. *Breast Cancer Res* 2007;9:R74 .
 52. Fehm T, Hoffmann O, Aktas B, Becker S, Solomayer EF, Wallwiener D, et al. Detection and characterization of circulating tumor cells in blood of primary breast cancer patients by RT-PCR and comparison to status of bone

Circulating tumor cells

- marrow disseminated cells. *Breast Cancer Res* 2009;11:R59.
53. Munzone E, Nole F, Goldhirsch A, Botteri E, Esposito A, Zorzino L, et al. Changes of HER2 status in circulating tumor cells compared with the primary tumor during treatment for advanced breast cancer. *Clin Breast Cancer* 2010;10:392-7 .
 54. Agelaki S, Kalykaki A, Markomanolaki H, Papadaki MA, Kallergi G, Hatzidaki D, et al. Efficacy of Lapatinib in Therapy-Resistant HER2-Positive Circulating Tumor Cells in Metastatic Breast Cancer. *PLoS ONE* 2015;10:e0123683.
 55. De Luca F, Rotunno G, Salvianti F, Galardi F, Pestrin M, Gabellini S, et al. Mutational analysis of single circulating tumor cells by next generation sequencing in metastatic breast cancer. *Oncotarget* 2016;7:26107-19.
 56. Reijm EA, Sieuwerts AM, Smid M, Vries JB, Mostert B, Onstenk W, et al. An 8-gene mRNA expression profile in circulating tumor cells predicts response to aromatase inhibitors in metastatic breast cancer patients. *BMC Cancer* 2016;16:123 .
 57. Pierga JY, Hajage D, Bachelot T, Delaloge S, Brain E, Campone M, et al. High independent prognostic and predictive value of circulating tumor cells compared with serum tumor markers in a large prospective trial in first-line chemotherapy for metastatic breast cancer patients. *Ann Oncol* 2012;23:618-24 .
 58. Hartkopf AD, Wagner P, Wallwiener D, Fehm T, Rothmund R. Changing levels of circulating tumor cells in monitoring chemotherapy response in patients with metastatic breast cancer. *Anticancer Res* 2011;31:979-84 .
 59. Muller V, Stahmann N, Riethdorf S, Rau T, Zabel T, Goetz A, et al. Circulating tumor cells in breast cancer: correlation to bone marrow micrometastases, heterogeneous response to systemic therapy and low proliferative activity. *Clin Cancer Res* 2005;11:3678-85 .
 60. Shao Y, Sun X, He Y, Liu C, Liu H. Elevated Levels of Serum Tumor Markers CEA and CA15-3 Are Prognostic Parameters for Different Molecular Subtypes of Breast Cancer. *Plos One* 2015;10:e0133830.
 61. Ebeling FG, Stieber P, Untch M, Nagel D, Konecny GE, Schmitt UM, et al. Serum CEA and CA 15-3 as prognostic factors in primary breast cancer. *Br J Cancer* 2002;86:1217-22.
 62. Konig A, Vilsmaier T, Rack B, Friese K, Janni W, Jeschke U, et al. Determination of Interleukin-4, -5, -6, -8 and -13 in Serum of Patients with Breast Cancer Before Treatment and its Correlation to Circulating Tumor Cells. *Anticancer Res* 2016;36:3123-30.
 63. Lu J, Fan T, Zhao Q, Zeng W, Zaslavsky E, Chen JJ, et al. Isolation of circulating epithelial and tumor progenitor cells with an invasive phenotype from breast cancer patients. *Int J Cancer* 2010;126:669-83 .
 64. Nakagawa T, Martinez SR, Goto Y, Koyanagi K, Kitago M, Shingai T, et al. Detection of circulating tumor cells in early-stage breast cancer metastasis to axillary lymph nodes. *Clin Cancer Res* 2007;13:4105-10.
 65. Mazel M, Jacot W, Pantel K, Bartkowiak K, Topart D, Cayrefourcq L, et al. Frequent expression of PD-L1 on circulating breast cancer cells. *Mol Oncol* 2015;9:1773-82 .
 66. Mikulova V, Cabinakova M, Janatkova I, Mestek O, Zima T, Tesarova P. Detection of circulating tumor cells during follow-up of patients with early breast cancer: Clinical utility for monitoring of therapy efficacy. *Scand J Clin Lab Invest* 2014;74:132-42 .
 67. Serrano MJ, Rovira PS, Martinez-Zubiaurre I, Rodriguez MD, Fernandez M, Lorente JA. Dynamics of circulating tumor cells in early breast cancer under neoadjuvant therapy. *Exp Ther Med* 2012;4:43-8.
 68. Soltani S, Mokarian F, Panjehpour M. The expression of CK-19 gene in circulating tumor cells of blood samples of metastatic breast cancer women. *Res Pharm Sci* 2015;10:485-96.
 69. Spiliotaki M, Mavroudis D, Kapranou K, Markomanolaki H, Kallergi G, Koinis F, et al. Evaluation of proliferation and apoptosis markers in circulating tumor cells of women with early breast cancer who are candidates for tumor dormancy. *Breast Cancer Res* 2014;16(6):485-96 .
 70. Guttery DS, Page K, Hills A, Woodley L, Marchese SD, Rghebi B, et al. Noninvasive detection of activating estrogen receptor 1 (ESR1) mutations in estrogen receptor-positive metastatic breast cancer. *Clin Chem* 2015;61:974-82 .
 71. Divella R, Daniele A, Savino E, Palma F, Bellizzi A, Giotta F, et al. Circulating levels of transforming growth factor-betaeta (TGF-beta) and chemokine (C-X-C motif) ligand-1 (CXCL1) as predictors of distant seeding of circulating tumor cells in patients with metastatic breast cancer. *Anticancer Res* 2013;33:1491-7.
 72. Stott SL, Hsu CH, Tsukrov DI, Yu M, Miyamoto DT, Waltman BA, et al. Isolation of circulating tumor cells using a microvortex-generating herringbone-chip. *Proc Natl Acad Sci U S A* 2010;107:18392-7 .
 73. Etayash H, Jiang K, Azmi S, Thundat T, Kaur K. Real-time Detection of Breast Cancer Cells Using Peptide-functionalized Microcantilever Arrays. *Sci Rep* 2015;5:13967.
 74. Mostert B, Sleijfer S, Foekens JA, Gratama JW. Circulating tumor cells (CTCs): detection methods and their clinical relevance in breast cancer. *Cancer Treat Rev* 2009;35:463-74 .

75. Wen CY, Wu LL, Zhang ZL, Liu YL, Wei SZ, Hu J, et al. Quick-response magnetic nanospheres for rapid, efficient capture and sensitive detection of circulating tumor cells. *ACS Nano* 2014;8:941-9.
76. Yu M, Stott S, Toner M, Maheswaran S, Haber DA. Circulating tumor cells: approaches to isolation and characterization. *J Cell Biol* 2011;192:373-82.
77. Hyun KA, Lee TY, Lee SH, Jung HI. Two-stage microfluidic chip for selective isolation of circulating tumor cells (CTCs). *Biosens Bioelectron* 2015;67:86-92.
78. Lim YC, Wiegman AP. Tracking metastatic breast cancer: the future of biology in biosensors. *Med Oncol* 2016;33:1-8.
79. Lim YC, Wiegman AP. Tracking metastatic breast cancer: the future of biology in biosensors. *Med Oncol* (Northwood, London, England) 2016;33:36 .
80. Litvinov SV, Velders MP, Bakker HA, Fleuren GJ, Warnaar SO. Ep-CAM: a human epithelial antigen is a homophilic cell-cell adhesion molecule. *J Cell Biol* 1994;125:437-46 .
81. Maetzel D, Denzel S, Mack B, Canis M, Went P, Benk M, et al. Nuclear signalling by tumour-associated antigen EpCAM. *Nat Cell Biol* 2009;11:162-71 .
82. Osta WA, Chen Y, Mikhitarian K, Mitas M, Salem M, Hannun YA, et al. EpCAM is overexpressed in breast cancer and is a potential target for breast cancer gene therapy. *Cancer Res* 2004;64:5818-24.
83. Schneck H, Gierke B, Uppenkamp F, Behrens B, Niederacher D, Stoecklein NH, et al. EpCAM-Independent Enrichment of Circulating Tumor Cells in Metastatic Breast Cancer. *Plos One* 2015;10:e0144535.
84. Mostert B, Kraan J, Sieuwerts AM, van der Spoel P, Bolt-de Vries J, Prager-van der Smissen WJ, et al. CD49f-based selection of circulating tumor cells (CTCs) improves detection across breast cancer subtypes. *Cancer Lett* 2012;319:49-55 .
85. Lu J, Fan T, Zhao Q, Zeng W, Zaslavsky E, Chen JJ, et al. Isolation of circulating epithelial and tumor progenitor cells with an invasive phenotype from breast cancer patients. *Int J Cancer* 2010;126:669-83.
86. Fan T, Zhao Q, Chen JJ, Chen WT, Pearl ML. Clinical significance of circulating tumor cells detected by an invasion assay in peripheral blood of patients with ovarian cancer. *Gynecol Oncol* 2009;112:185-91 .
87. Paris PL, Kobayashi Y, Zhao Q, Zeng W, Sridharan S, Fan T, et al. Functional phenotyping and genotyping of circulating tumor cells from patients with castration resistant prostate cancer. *Cancer Lett* 2009;277:164-73.
88. Bidard F-C, Peeters DJ, Fehm T, Nolé F, Gisbert-Criado R, Mavroudis D, et al. Clinical validity of circulating tumour cells in patients with metastatic breast cancer: a pooled analysis of individual patient data. *Lancet Oncol* 2014;15:406-14.
89. Dawood S, Broglio K, Valero V, Reuben J, Handy B, Islam R, et al. Circulating tumor cells in metastatic breast cancer: from prognostic stratification to modification of the staging system? *Cancer* 2008;113:2422-30 .
90. Franken B, de Groot MR, Mastboom WJ, Vermes I, van der Palen J, Tibbe AG, et al. Circulating tumor cells, disease recurrence and survival in newly diagnosed breast cancer. *Breast Cancer Res* 2012;14:1-8.
91. Hayes DF, Cristofanilli M, Budd GT, Ellis MJ, Stopeck A, Miller MC, et al. Circulating tumor cells at each follow-up time point during therapy of metastatic breast cancer patients predict progression-free and overall survival. *Clin Cancer Res* 2006;12:4218-24 .
92. Nole F, Munzone E, Zorzino L, Minchella I, Salvatici M, Botteri E, et al. Variation of circulating tumor cell levels during treatment of metastatic breast cancer: prognostic and therapeutic implications. *Ann Oncol* 2008;19:891-7 .
93. Wallwiener M, Riethdorf S, Hartkopf AD, Modugno C, Nees J, Madhavan D, et al. Serial enumeration of circulating tumor cells predicts treatment response and prognosis in metastatic breast cancer: a prospective study in 393 patients. *BMC Cancer* 2014;14:1-12.
94. Lowe AC, Pignon JC, Carvo I, Drage MG, Constantine NM, Jones N, et al. Young investigator challenge: Application of cytologic techniques to circulating tumor cell specimens: Detecting activation of the oncogenic transcription factor STAT3. *Cancer Cytopathol* 2015;123:696-706.
95. Forte VA, Barrak DK, Elhodaky M, Tung L, Snow A, Lang JE. The potential for liquid biopsies in the precision medical treatment of breast cancer. *Cancer Biol Med* 2016;13:19-40.
96. Schramm A, Friedl TW, Schochter F, Scholz C, de Gregorio N, Huober J, et al. Therapeutic intervention based on circulating tumor cell phenotype in metastatic breast cancer: concept of the DETECT study program. *Arch Gynecol Obstet* 2016;293:271-81.
97. Sigal EK, Jeffrey SS, eds. *Tumor Heterogeneity and Single-cell Analysis of CTCs. Circulating Tumor Cells: John Wiley & Sons, 2016:313-28.*
98. Tewes M, Aktas B, Welt A, Mueller S, Hauch S, Kimmig R, et al. Molecular profiling and predictive value of circulating tumor cells in patients with metastatic breast cancer: an option for monitoring response to breast cancer related therapies. *Breast Cancer Res Treat* 2009;115:581-90.
99. Fehm T, Muller V, Aktas B, Janni W, Schneeweiss A, Stickeler E, et al. HER2 status of circulating tumor cells in patients with metastatic breast cancer: a prospective,

Circulating tumor cells

- multicenter trial. *Breast Cancer Res Treat* 2010;124:403-12.
100. Park YH, Shin HT, Jung HH, Choi YL, Ahn T, Park K, et al. Role of HER2 mutations in refractory metastatic breast cancers: targeted sequencing results in patients with refractory breast cancer. *Oncotarget* 2015;6:32027-38 .
 101. Allwien M, Hartkopf AD, Baccelli I, Riethdorf S, Schott S, Pantel K, et al. The prognostic impact of circulating tumor cells in subtypes of metastatic breast cancer. *Breast Cancer Res Treat* 2013;137:503-10 .
 102. Wulfing P, Borchard J, Buerger H, Heidl S, Zanker KS, Kiesel L, et al. HER2-positive circulating tumor cells indicate poor clinical outcome in stage I to III breast cancer patients. *Clin Cancer Res* 2006;12:1715-20 .
 103. Giuliano M, Giordano A, Jackson S, Hess KR, De Giorgi U, Mego M, et al. Circulating tumor cells as prognostic and predictive markers in metastatic breast cancer patients receiving first-line systemic treatment. *Breast Cancer Res* 2011;13:67.
 104. Gentile A, Trusolino L, Comoglio PM. The Met tyrosine kinase receptor in development and cancer. *Cancer Metastasis Rev* 2008;27:85-94 .
 105. Gires O, Klein CA, Baeuerle PA. On the abundance of EpCAM on cancer stem cells. *Nat Rev Cancer* 2009;9:143.
 106. Shimada K, Nakajima A, Ikeda K, Ishibashi K, Shimizu N, Ito K. CD47 regulates the TGF-beta signaling pathway in osteoblasts and is distributed in Meckel's cartilage. *J Oral Sci* 2011;53:169-75.
 107. Ascolani G, Occhipinti A, Liò P. Modelling Circulating Tumour Cells for Personalised Survival Prediction in Metastatic Breast Cancer. *PLoS Comput Biol* 2015;11:e1004199.
 108. Roy-Chowdhuri S, de Melo Gagliato D, Routbort MJ, Patel KP, Singh RR, Broaddus R, et al. Multigene Clinical Mutational Profiling of Breast Carcinoma Using Next-Generation Sequencing. *American J Clin Pathol* 2015;144:713-21.
 109. Fuhrmann C, Schmidt-Kittler O, Stoecklein NH, Petat-Dutter K, Vay C, Bockler K, et al. High-resolution array comparative genomic hybridization of single micrometastatic tumor cells. *Nucleic Acids Res* 2008;36:e39-e.
 110. Stoecklein NH, Hosch SB, Bezler M, Stern F, Hartmann CH, Vay C, et al. Direct Genetic Analysis of Single Disseminated Cancer Cells for Prediction of Outcome and Therapy Selection in Esophageal Cancer. *Cancer Cell* 2008;13:441-53.
 111. Navin N, Kendall J, Troge J, Andrews P, Rodgers L, McIndoo J, et al. Tumour evolution inferred by single-cell sequencing. *Nature*. 2011;472:90-4.
 112. Heitzer E, Auer M, Gasch C, Pichler M, Ulz P, Hoffmann EM, et al. Complex tumor genomes inferred from single circulating tumor cells by array-CGH and next-generation sequencing. *Cancer Res* 2013;73:2965-75 .
 113. Pestrin M, Salvianti F, Galardi F, De Luca F, Turner N, Malorni L, et al. Heterogeneity of PIK3CA mutational status at the single cell level in circulating tumor cells from metastatic breast cancer patients. *Mol Oncol* 2015;9:749-57.
 114. Polzer B, Medoro G, Pasch S, Fontana F, Zorzino L, Pestka A, et al. Molecular profiling of single circulating tumor cells with diagnostic intention. *EMBO Mol Med* 2014;6:1371-86.
 115. Yu M, Bardia A, Aceto N, Bersani F, Madden MW, Donaldson MC, et al. Ex vivo culture of circulating breast tumor cells for individualized testing of drug susceptibility. *Science* 2014;345:216-20.
 116. Helzer KT, Barnes HE, Day L, Harvey J, Billings PR, Forsyth A. Circulating tumor cells are transcriptionally similar to the primary tumor in a murine prostate model. *Cancer Res* 2009;69:7860-6 .
 117. Marchetti A, Del Gramastro M, Felicioni L, Malatesta S, Filice G, Centi I, et al. Assessment of *EGFR* Mutations in Circulating Tumor Cell Preparations from NSCLC Patients by Next Generation Sequencing: Toward a Real-Time Liquid Biopsy for Treatment. *PLoS ONE* 2014;9:e103883.
 118. Yu M, Bardia A, Aceto N, Bersani F, Madden MW, Donaldson MC, et al. Ex vivo culture of circulating breast tumor cells for individualized testing of drug susceptibility. *Science (New York, NY)* 2014;345:216-20.
 119. Cristofanilli M. Circulating tumor cells, disease progression, and survival in metastatic breast cancer. *Semin Oncol* 2006;33:S9-14.
 120. Hayes DF, Cristofanilli M, Budd GT, Ellis MJ, Stopeck A, Miller MC, et al. Circulating tumor cells at each follow-up time point during therapy of metastatic breast cancer patients predict progression-free and overall survival. *Clin Cancer Res* 2006;12:4218-24.
 121. Liu MC, Shields PG, Warren RD, Cohen P, Wilkinson M, Ottaviano YL, et al. Circulating tumor cells: a useful predictor of treatment efficacy in metastatic breast cancer. *J Clin Oncol* 2009;27:5153-9.
 122. Lucci A, Hall CS, Lodhi AK, Bhattacharyya A, Anderson AE, Xiao LC, et al. Circulating tumour cells in non-metastatic breast cancer: a prospective study. *Lancet Oncol* 2012;13:688-95.
 123. Nakamura S, Yagata H, Ohno S, Yamaguchi H, Iwata H, Tsunoda N, et al. Multi-center study evaluating circulating tumor cells as a surrogate for response to

- treatment and overall survival in metastatic breast cancer. *Breast Cancer* 2010;17:199-204.
124. Pachmann K, Camara O, Kavallaris A, Krauspe S, Malarski N, Gajda M, et al. Monitoring the response of circulating epithelial tumor cells to adjuvant chemotherapy in breast cancer allows detection of patients at risk of early relapse. *J Clin Oncol* 2008;26:1208-15.
 125. Rack B, Schindlbeck C, Jückstock J, Andergassen U, Hepp P, Zwingers T, et al. Circulating Tumor Cells Predict Survival in Early Average-to-High Risk Breast Cancer Patients. *J Natl Cancer Inst* 2014;106:dju066.
 126. Satelli A, Brownlee Z, Mitra A, Meng QH, Li S. Circulating tumor cell enumeration with a combination of epithelial cell adhesion molecule- and cell-surface vimentin-based methods for monitoring breast cancer therapeutic response. *Clin Chem* 2015;61:259-66.
 127. Serrano MJ, Sanchez-Rovira P, Delgado-Rodriguez M, Gaforio JJ. Detection of circulating tumor cells in the context of treatment: prognostic value in breast cancer. *Cancer Biol Ther* 2009;8:671-5.
 128. de Albuquerque A, Kaul S, Breier G, Krabisch P, Fersis N. Multimarker Analysis of Circulating Tumor Cells in Peripheral Blood of Metastatic Breast Cancer Patients: A Step Forward in Personalized Medicine. *Breast care (Basel, Switzerland)* 2012;7:7-12.
 129. Gradilone A, Naso G, Raimondi C, Cortesi E, Gandini O, Vincenzi B, et al. Circulating tumor cells (CTCs) in metastatic breast cancer (MBC): prognosis, drug resistance and phenotypic characterization. *Ann Oncol* 2010;22:86-92.
 130. Gradilone A, Raimondi C, Naso G, Silvestri I, Repetto L, Palazzo A, et al. How circulating tumor cells escape from multidrug resistance: translating molecular mechanisms in metastatic breast cancer treatment. *Am J Clin Oncol* 2011;34:625-7.
 131. Jansson S, Bendahl P-O, Larsson A-M, Aaltonen KE, Rydén L. Prognostic impact of circulating tumor cell apoptosis and clusters in serial blood samples from patients with metastatic breast cancer in a prospective observational cohort. *BMC Cancer* 2016;16:433.
 132. Reinholz MM, Kitzmann KA, Tenner K, Hillman D, Dueck AC, Hobday TJ, et al. Cytokeratin-19 and mammaglobin gene expression in circulating tumor cells from metastatic breast cancer patients enrolled in North Central Cancer Treatment Group trials, N0234/336/436/437. *Clin Cancer Res* 2011;17:7183-93.
 133. Tewes M, Aktas B, Welt A, Mueller S, Hauch S, Kimmig R, et al. Molecular profiling and predictive value of circulating tumor cells in patients with metastatic breast cancer: an option for monitoring response to breast cancer related therapies. *Breast Cancer Res Treat* 2009;115:581-90.
 134. Wallwiener M, Hartkopf AD, Baccelli I, Riethdorf S, Schott S, Pantel K, et al. The prognostic impact of circulating tumor cells in subtypes of metastatic breast cancer. *Breast Cancer Res Treat* 2013;137:503-10.
 135. Wang HY, Ahn S, Kim S, Park S, Han H, Sohn JH, et al. Detection of circulating tumor cells in patients with breast cancer using the quantitative RT-PCR assay for monitoring of therapy efficacy. *Exp Mol Pathol* 2014;97:445-52.
 136. Zehentner BK, Secrist H, Hayes DC, Zhang X, Ostenson RC, Loop S, et al. Detection of circulating tumor cells in peripheral blood of breast cancer patients during or after therapy using a multigene real-time RT-PCR assay. *Molecular diagnosis Ther* 2006;10:41-7.