

# Regulatory Functions of MicroRNAs in Cancer Pathogenesis

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**Abstract-** MicroRNAs (miRNAs) are a large family of evolutionary conserved small non-coding RNA molecules that firstly discovered in 1993. They regulate gene expression of about 50% of protein-coding genes at the post-transcriptional level. MiRNAs can target numerous messenger RNAs and subsequent misexpression of them can affect many different signaling pathways. They are playing a pivotal role in cancer development by regulation of the genes expression which involved in the proliferation, survival, differentiation, apoptosis or metastasis of the cancer cells. Several treatment approaches such as inhibition of oncomiRs and restoration of tumor suppressor miRNAs have been established in certain types of cancers and some other miRNA-based strategies are in development for cancer prevention and treatment. Nowadays, cancer is the most important target of miRNA therapeutics and the specific mechanisms by which miRNA mediates cancer pathways needs more research and study.

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## Introduction

MicroRNAs (miRNAs) are a large family of evolutionary conserved single-stranded small RNA molecules which occur as non-coding RNAs of 19-24 nucleotides in length (1,2). MiRNAs regulate gene expression of about 50% of any protein-coding gene at the posttranscriptional level (3). In the posttranscriptional level, miRNAs act through the degradation of their target mRNA or translational inhibition of the entitled mRNA (4). MiRNA coding genes are located either in intergenic regions or in the introns and are transcribed by RNA polymerase II into long primary transcripts called primary miRNAs (pri-miRNAs) (5). In the nucleus, these are processed by RNAase III endonuclease Drosha and double-stranded RNA-binding protein Pasha, into a structure called precursor-miRNA (pre-miRNA) (6). Pre-miRNAs are transported to the cytoplasm. There, they are cleaved, and a short RNA duplex molecule is generated (7). Later a helicase forms the mature miRNA. Mature miRNA is then assembled into the RNA induced silencing complex (RISC) (8,9). RISC regulates target mRNA's function by binding to it and silencing its expression (10-12). In

addition, by acting on regulatory sequences of their target gene, miRNAs can promote the expression (13). MiRNAs mostly can target numerous mRNAs, thus in case of misexpression in a single miRNA, expression of several hundreds of proteins can be disrupted, and many different signaling pathways may be affected. This also can cause cancerous transformation (14). The processing of miRNAs is demonstrated in Figure 1.

## MicroRNA discovery

MicroRNA was first discovered by Victor Ambros' laboratory in 1993 during research on *Caenorhabditis elegans*. Simultaneously, Gary Ravkun reported the first miRNA target gene, which resulted in the identification of a novel mechanism of posttranscriptional gene regulation (15). Later, Ravukon and Horvitz found let-7 in the same model nematode species. Also, a class of short (small) interfering RNA (siRNA) involved in the process of RNA interference was discovered. Following these findings, the various number of miRNAs have been discovered and reported in mammals, and more than 700 miRNAs, which were identified in humans, have been fully sequenced (16).

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## MicroRNA and cancer

Proliferation, Differentiation, development, and metabolism are examples of multiple biological functions of this class RNAs. In addition, miRNAs are playing an important role in cancer, diabetes, autism, fragile X syndrome, Alzheimer's, and heart diseases (17-20). Cancer has been known as a common complex disease worldwide. Series of genetic and epigenetic factors alter certain balances, which cause uncontrolled cellular proliferation. Due to the complexity of cancer, a single therapeutic strategy will not be able to produce a lasting cure. MiRNAs, however, got the capacity to target several protein-coding genes at the same time. As a result, a small change in miRNA expression can lead to meaningful alterations in the expression profile of several protein-coding genes and, therefore, cause changes in cellular phenotype (21-23). MiRNAs can be classified as oncomiR and tumor suppressor miRNAs from which a large number of can be used as diagnostic and prognostic biomarkers of the cancers (24,25).

Tumor formation is the result of alterations in miRNA expression by decreasing the expression of necessary genes that are needed in the proliferation or survival of the cells. However, in another study, it has been indicated that cancer progression or tumorigenesis does not contribute directly to miRNA (26). It is not completely found out that the changes in miRNA expression are either because of the pathological state of cancer or the cancer is the direct reason for it. Nevertheless, miRNA expression is affected directly or indirectly by several alterations that happen in cancer cells. Some changes such as gene mutations, changes in epigenetic regulation of miRNA, abnormalities in miRNA genes, or proteins that are involved in their construction and genomic rearrangements are some of the examples of alterations that might affect miRNA expression.

One of the main factors of changing miRNA expression in tumor cells in the presence of miRNAs in tumor-related genomic regions or fragile genomic areas. This specifically causes influences miRNA and mRNA connectivity features, which can be named the direct effect of the mutations. Incomplete translational processes are the result of altered miRNA interactions (27). MiRNAs are capable of regulating a broad set of genes efficiently and silence target genes simultaneously. Since cancer is a heterogenic disease, miRNA's characteristic features are beneficial for treatment. MiRNAs target cancer cells in spite of targeting endothelial cells and fibroblasts. This helps the inhibiting of angiogenesis and tumor fibrosis. Therefore,

the required process during metastasis and tumor formation is blocked (28). Moreover, dysregulated miRNAs are implicated in the pathogenesis of cancer due to having an effect on oncogenes and/or tumor suppressor genes (29,30).

## MicroRNAs function

The role of miRNA in cancer has already been intensively evaluated, and either clinical studies or in vitro and in vivo experiments demonstrated their importance on this occasion (23). MiRNA's role was first studied in association with chronic lymphocytic leukemia (CLL) (31). Later multiple miRNAs were reported in accordance with plenty of other cancers (32-40). The dysregulation of miRNAs is also linked to cancer in various studies (41-44). It has been proved that they play important roles in metastasis, initiation, and progression, as well as therapeutic resistance (44). There also exists researches describing miRNAs playing two separate acts in carcinogenesis (to be both as "oncomirs" and as "tumor suppressors") (45-48). In support of this thought, scientists demonstrated the fact that miRNA expression can be up- or down-regulated in cancer cells in comparison with normal cells. They also seem to be deregulated in hematological malignancies, as well as many solid tumors (42,49,50). When it comes to location, however, about 50% of miRNA genes are embedded in genomic instability regions (51). This strengthens the evidence of cancer being related to miRNAs. Besides, miRNAs regulate 20-30% of all protein-coding genes (52,53), which supports the probability of miRNA's signature, providing efficient information about tumors (49).

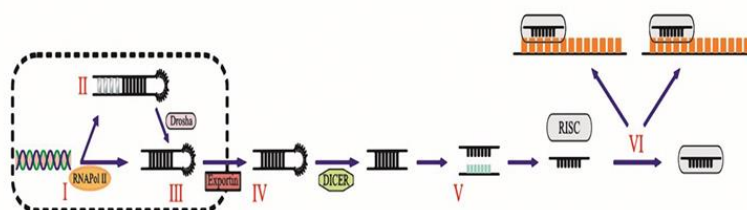
Notably, expression patterns of miRNAs are tumor- and tissue-specific (32,54,55). For example, miR-155 is a multifunctional miRNA and is involved in inflammation, immune response, and cancer development (56), over-expressed in leukemia and lymphoma (57-60), and down-regulated in melanoma, gastric cancer, ovarian cancer, and endocrine tumors (61-65). These capabilities make miRNAs valuable agents for diagnosis and therapy of certain kinds of cancer. Another example of cancer-specific miRNAs is miR-21, which has been investigated by several groups. Subsequently, scientists found out that miR-21 is over-expressed in malignancies like a breast (50), colon (66), lung (32), liver (67), thyroid (68), and leukemia (32). These findings suggested that this oncomir is a good example of a cancer-specific miRNA (69).

Dysfunction or misexpression of miRNA can affect a broad range of processes involved in tumor progressions

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such as metastasis, apoptosis, angiogenesis, and cell cycle regulation (70-75). There are reports of five families of miRNA who target cell cycle regulators. These miRNAs are the let-7, the miR-15a/16 cluster, miR-34 families, the miR-17/20 cluster, and the miR-221/222 cluster. The entitled miRNAs are capable of controlling cell cycle checkpoints. Malfunction of these miRNAs may cause a rise in cell proliferation, which is necessary for tumor growth (76). However, some studies demonstrated miRNAs as anti-apoptotic regulators of key pathways in cancer. These miRNAs are highlighted

to maintain cancer cell survival and drug resistance contribution (77). Besides, pro-apoptotic miRNAs serving as anti-cancer agents (78). The miRNAs also play a key role in the metabolism of cancer cells (79). They regulate nutrient uptake, targeting transporters, and metabolic enzymes and modulating cancer cell metabolism. They increase the accumulation of materials to control metabolic flux and support proliferation (80,81). We summarized some important onco-miRNAs and their functions in Table 1.



**Figure 1.** MicroRNA processing. RNA polymerase II and proper transcription factors excite the transcription of the microRNA gene (I) in the production of a pri-miRNA. The primary transcript (II) is then processed by an RNAase III enzyme called Drosha to produce a ~ 65 nucleotide (nt) pre-miRNA. The pre-miRNA, which has a short stem of 2–3 nt 3' overhangs (III), is then exported by exportin 5 (EXP5) to the cytoplasm for additional processing. In the cytoplasm, the precursor microRNA subsequently processed into a mature 19–24 nucleotide duplex (IV) by an enzyme called Dicer. Afterward, the duplex is separated into a primary and secondary strand (V); then, the primary strand is embedded into the RISC (RNA-induced silencing complex). In the next step, the microRNA with RISC targets complementary mRNA transcripts (VI) at the seed region to induce either block translation (right) or mRNA degradation (left)

**Table 1. Some important onco-miRNA and their function**

MicroRNAS	Target gene(S) or Protein	Function
<b>miR 15 and miR 16</b>	NA	B-cell lymphocytic, chronic Leukemia
<b>miR-17 ~ 92 cluster</b>	NA	Lung and other malignancies
	Myc	Tumorigenesis and angiogenesis
<b>miR-21</b>	Pcd4, BMPRII & LRRFIP1	Promote apoptosis through activation of caspases
<b>miR-155</b>	TP53INP1	Overexpression in pancreatic cancer and breast cancer progression
<b>miR-371 ~ 3</b>	LATS2	Cell proliferation and tumor development

NA: Not applicable

Metastasis-mediating miRNAs have also been discovered. They regulate distinct steps of the metastasis, affecting both signaling pathways in the cancer cell and interactions of cells with one another and with tumor stroma. According to studies, they can activate or suppress metastasis (82,83). Some miRNAs are implicated in suppressing apoptosis and stimulating tumorigenesis (84). Other miRNA families contribute to both tumor growth and metastasis (38). They are able to silence multiple oncogenes and are down-regulated in several tumors (85,86). Pro-metastatic miRNAs, however, are another example of metastasis-promoting miRNAs (87,88). Subsequently, due to the important

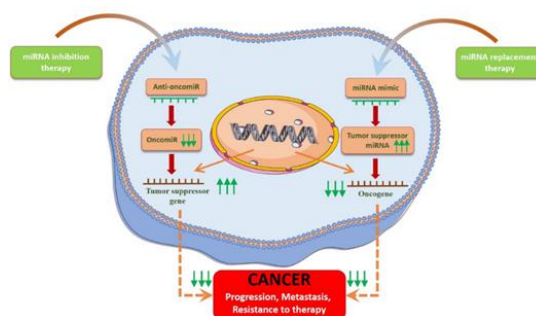
role of miRNAs in cancer, there exists a wide range of strategies based on miRNA in oncology. They can be used for cancer classification (49,89) or tissue origin identification of cancers with the unknown primary origin (90,91). Their expression can serve as a useful prognostic or diagnostic marker (92-95). Interestingly, miRNA signatures have been established as predictive factors of response to therapy (96-100) and drugs (101,102).

### MicroRNA in cancer therapy

Since miRNAs discovery, a debate has risen that miRNAs could be regarded as a promising biomarker to

improve response to cancer treatment (26). The advent of miRNA-based therapy, however, has opened new avenues to use targeted therapy for clinical applications, since there are some limitations for current cancer therapies (103). All of the applications in the previous section are possible when dealing with primary tumors. The fact that miRNAs are more stable than mRNAs is the key point since this stability enables them to be detected in the circulation and serve as biomarkers.

Circulating miRNAs can be measured with regard to a wide variety of cancers (104). Therefore, studies are currently highlighting their employment in cancer therapeutics (105). Based on several studies in recent years, miRNAs can be used as highly potential molecules in CRC therapy (30). Several therapeutic strategies associated with miRNAs can prevent cancer progression (Figure 2).



**Figure 2.** Different miRNA-based therapeutic strategies against cancer progression, metastasis, and resistance to therapy

Some examples include cutting oncogenic miRNAs by artificial miRNAs, which are capable of pairing with mRNAs, inhibition of the entitled oncogenic miRNAs, inducing the tumor suppressor miRNAs, or decreasing miRNA expression using various epigenetic factors like promoter methylation. To reduce the miRNA expression, antisense oligonucleotides can also be used. They are particularly paired with miRNA (26). One type that is artificially made is Antagomir (106). In comparison with other cancer treatment methods, these molecules are less toxic and create stable inhibition (27).

MiRNAs can also be agents or targets of cancer therapy according to their function, stage of cancer, and type of cancer (107). In order to use combined therapies targeting multiple miRNAs, tumor-secreted miRNA, who are messengers and/or effectors, must be characterized as the first step. Due to the correlation between their levels and metastasis, circulating or exosomal miRNAs can be quantified to select patients with high risks of metastasis in a certain type of cancer. As a result, these patients will benefit from a preventive strategy that targets the miRNA effectors (108). There also have been studies indicating that treatment interventions such as inhibition of oncomiRs and restoration of tumor suppressor miRNAs might be beneficial for certain types of cancers (25).

In addition to the fact that cell-free miRNAs are functionally effective in metastatic progression, they are also nominated for potential novel therapeutic targets

(108,109). An interesting advantage of cell-free miRNAs is that their expression levels can be monitored when treatment is started (110). In this regard circulating miRNAs are discovered to be potential diagnostic and therapeutic agents in association with cancers (111). Another aspect of targeting miRNAs is that they are observed to be beneficial for improved response to drugs. Hence, circulating miRNA's expression level in blood is useful for prognosis determination (102,112). Moreover, compared to other gene-therapy methods and drug molecules, miRNA showed low toxicity (101). Accordingly, in case of safe delivery to cancer cells, miRNA-based therapeutics seems to be promising anti-cancer guardians.

Once miRNAs discovered, significant progress in the identification of these novel family has confirmed that these small and non-coding RNAs are a numerous class of regulatory RNAs. Also, the skeleton of a biochemical mechanism for their functions in gene regulation has specified. The most attractive part of miRNA therapeutics is their capability to target any genes, which is not possible or difficult by protein-based drugs or small molecules. Nowadays, cancer is the most important target of miRNA therapeutics among the numerous diseases being studied. We briefly clarified the particular roles and the importance of miRNAs in the regulation of gene expression. Additionally, the specific mechanisms by which miRNA mediated repression needs more research and study.

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## References

1. Ling H, Zhang W, Calin GA. Principles of microRNA involvement in human cancers. *Chin J Cancer* 2011;30:739-48 .
2. Mattick JS, Makunin I V. Non-coding RNA. *Hum Mol Genet* 2006;15:17-29 .
3. Mirakholi M, Mahmoudi T, Heidari M. MicroRNAs Horizon in Retinoblastoma. *Acta Med Iran.* 51(12):823-829.
4. Bartel DP. MicroRNAs: genomics, biogenesis, mechanism, and function. *Cell* 2004;116:281-97.
5. Lee Y, Jeon K, Lee JT, Kim S, Kim VN. MicroRNA maturation: Stepwise processing and subcellular localization. *EMBO J* 2002;21:4663-70 .
6. Han J, Lee Y, Yeom K-H, Kim Y-K, Jin H, Kim VN. The Drosha-DGCR8 complex in primary microRNA processing. *Genes Dev* 2004;18:3016-27 .
7. Yi R, Qin Y, Macara IG, Cullen BR. Exportin-5 mediates the nuclear export of pre-microRNAs and short hairpin RNAs. *Genes Dev* 2003;17:3011-6.
8. Lewis BP, Burge CB, Bartel DP. Conserved seed pairing, often flanked by adenosines, indicates that thousands of human genes are microRNA targets. *Cell* 2005;120:15-20 .
9. Denli AM, Tops BBJ, Plasterk RHA, Ketting RF, Hannon GJ. Processing of primary microRNAs by the Microprocessor complex. *Nature* 2004;432:231-5.
10. Bartel DP. MicroRNAs: Target Recognition and Regulatory Functions. *Cell* 2009;136:215-33.
11. Siomi H, Siomi MC. Posttranscriptional Regulation of MicroRNA Biogenesis in Animals. *Mol Cell* 2010;38:323-32.
12. Bhayani MK, Calin GA, Lai SY. Functional relevance of miRNA\* sequences in human disease. *Mutat Res* 2012;731:14-9.
13. Wiemer EAC. The role of microRNAs in cancer: No small matter. *Eur J Cancer* 2007;43:1529-44.
14. Jansson MD, Lund AH. MicroRNA and cancer. *Mol Oncol* 2012;6:590-610 .
15. Ambros V. The functions of animal microRNAs. *Nature* 2004;431:350-5.
16. Mohammadi A, Mansoori B, Baradaran B. Regulation of miRNAs by herbal medicine: An emerging field in cancer therapies. *Biomed Pharmacother* 2017;86:262-70.
17. Le Quesne J, Caldas C. Micro-RNAs and breast cancer. *Mol Oncol* 2010;4:230-41.
18. Almeida MI, Reis RM, Calin GA. MicroRNA history: Discovery, recent applications, and next frontiers. *Mutat Res* 2011;717:1-8.
19. Shenouda SK, Alahari SK. MicroRNA function in cancer: Oncogene or a tumor suppressor? *Cancer Metastasis Rev* 2009;28:369-78.
20. Huang Y, Shen XJ, Zou Q, Wang SP, Tang SM, Zhang GZ. Biological functions of microRNAs: A review. *J Physiol Biochem* 2011;67:129-39.
21. Mohammadzadeh R, Baradaran B, Valizadeh H, Yousefi B, Zakeri-Milani P. Reduced ABCB1 Expression and Activity in the Presence of Acrylic Copolymers. *Adv Pharm Bull.* 2014;4:219-24.
22. Suárez Y, Sessa WC. MicroRNAs As Novel Regulators of Angiogenesis. *Circ Res* 2009;104:442-54.
23. Spizzo R, Nicoloso MS, Croce CM, Calin GA. SnapShot: MicroRNAs in Cancer. *Cell* 2009;137:586.e1.
24. Wang JY, Huang JC, Chen G, Wei DM. Expression level and potential target pathways of miR-1-3p in colorectal carcinoma based on 645 cases from 9 microarray datasets. *Mol Med Rep* 2018;17:5013-20.
25. Karimi Dermani F, Azizi Jalilian F, Hossienkhani H, Ezati R, Amini R. siRNA Delivery Technology for Cancer Therapy: Promise and Challenges. *Acta Med Iran.* 57(2):83-93.
26. Shirjang S, Mansoori B, Asghari S, Duijf PHG, Mohammadi A, Gjerstorff M, et al. MicroRNAs in cancer cell death pathways: Apoptosis and necroptosis. *Free Radic Biol Med.* 2019;139:1-15.
27. Schaefer A, Jung M, Kristiansen G, Lein M, Schrader M, Miller K, et al. MicroRNAs and cancer: Current state and future perspectives in urologic oncology. *Urol Oncol* 2010;28:4-13.
28. Plummer PN, Freeman R, Taft RJ, Vider J, Sax M, Umer BA, et al. MicroRNAs regulate tumor angiogenesis modulated by endothelial progenitor cells. *Cancer Res* 2013;73:341-52.
29. Mansoori B, Mohammadi A, Ghasabi M, Shirjang S, Dehghan R, Montazeri V, et al. miR- 142- 3p as tumor suppressor miRNA in the regulation of tumorigenicity, invasion and migration of human breast cancer by targeting Bach- 1 expression. *J Cell Physiol.* 2019;234:9816–25.
30. Slaby O, Svoboda M, Michalek J, Vyzula R. MicroRNAs in colorectal cancer: Translation of molecular biology into clinical application. *Mol Cancer* 2009;8:102.
31. Calin GA, Dumitru CD, Shimizu M, Bichi R, Zupo S, Noch E, et al. Nonlinear partial differential equations and applications: Frequent deletions and down-regulation of

- micro- RNA genes miR15 and miR16 at 13q14 in chronic lymphocytic leukemia. *Proc Natl Acad Sci U S A*. 2002;99:15524-9 .
32. Moazeni M, Khoramizadeh MR, Teimoori-Toolabi L, Noorbakhsh F, Rezaie S. The Effect of EFG1 Gene Silencing on Down-Regulation of SAP5 Gene, by Use of RNAi Technology. *Acta Med Iran*;52:9-14.
  33. Calin GA, Cimmino A, Fabbri M, Ferracin M, Wojcik SE, Shimizu M, et al. MiR-15a and miR-16-1 cluster functions in human leukemia. *Proc Natl Acad Sci U S A*. 2008;105:5166-71.
  34. Deng M, Tang H, Zhou Y, Zhou M, Xiong W, Zheng Y, et al. Mir-216b suppresses tumor growth and invasion by targeting KRAS in nasopharyngeal carcinoma. *J Cell Sci* 2011;124:2997-3005 .
  35. Díaz R, Silva J, García JM, Lorenzo Y, García V, Peña C, et al. Deregulated expression of miR-106a predicts survival in human colon cancer patients. *Genes Chromosomes Cancer* 2008;47:794-802.
  36. Drakaki A, Iliopoulos D. MicroRNA-gene signaling pathways in pancreatic cancer. *Biomed J* 2013;3:200-8.
  37. Hirata H, Ueno K, Shahryari V, Tanaka Y, Tabatabai ZL, Hinoda Y, et al. Oncogenic miRNA-182-5p Targets Smad4 and RECK in Human Bladder Cancer. *PLoS One* 2012;7:1-8.
  38. Takamizawa J, Konishi H, Yanagisawa K, Tomida S, Osada H, Endoh H, et al. Reduced expression of the let-7 microRNAs in human lung cancers in association with shortened postoperative survival. *Cancer Res* 2004;64:3753-6.
  39. Macha M, Seshacharyulu P, Krishn S, Pai P, Rachagani S, Jain M, et al. MicroRNAs (miRNAs) as Biomarker(s) for Prognosis and Diagnosis of Gastrointestinal (GI) Cancers. *Curr Pharm Des* 2014;20:5287-97.
  40. Radhakrishnan P, Mohr AM, Grandgenett PM, Steele MM, Batra SK, Hollingsworth MA. MicroRNA-200c Modulates the Expression of MUC4 and MUC16 by Directly Targeting Their Coding Sequences in Human Pancreatic Cancer. *PLoS One* 2013;8:e73356.
  41. Lee YS, Dutta A. MicroRNAs in Cancer. *Annu Rev Pathol* 2009;4:199-227.
  42. Garzon R, Calin GA, Croce CM. MicroRNAs in Cancer. *Annu Rev Med* 2009;60:167-79.
  43. Meltzer PS. Small RNAs with big impacts. *Nature* 2005;435:745-6.
  44. Calin GA, Croce CM. MicroRNA signatures in human cancers. *Nat Rev Cancer* 2006;6:857-66.
  45. Welch C, Chen Y, Stallings RL. MicroRNA-34a functions as a potential tumor suppressor by inducing apoptosis in neuroblastoma cells. *Oncogene* 2007;26:5017-22 .
  46. He L, Thomson JM, Hemann MT, Hernando-Monge E, Mu D, Goodson S, et al. A microRNA polycistron as a potential human oncogene. *Nature*. 2005;435:828-33.
  47. Yu S, Lu Z, Liu C, Meng Y, Ma Y, Zhao W, et al. miRNA-96 suppresses KRAS and functions as a tumor suppressor gene in pancreatic cancer. *Cancer Res* 2010;70:6015-25.
  48. Esquela-Kerscher A, Slack FJ. Oncomirs - MicroRNAs with a role in cancer. *Nat Rev Cancer* 2006;6:259-69.
  49. Lu J, Getz G, Miska EA, Alvarez-Saavedra E, Lamb J, Peck D, et al. MicroRNA expression profiles classify human cancers. *Nature* 2005;435:834-8.
  50. Iorio MV, Ferracin M, Liu CG, Veronese A, Spizzo R, Sabbioni S, et al. MicroRNA gene expression deregulation in human breast cancer. *Cancer Res* 2005;65:7065-70.
  51. Calin GA, Sevignani C, Dumitru CD, Hyslop T, Noch E, Yendamuri S, et al. Human microRNA genes are frequently located at fragile sites and genomic regions involved in cancers. *Proc Natl Acad Sci* 2004;101:2999-3004 .
  52. Bentwich I, Avniel A, Karov Y, Aharonov R, Gilad S, Barad O, et al. Identification of hundreds of conserved and nonconserved human microRNAs. *Nat Genet* 2005;37:766-70 .
  53. Carthew RW. Gene regulation by microRNAs. *Curr Opin Genet Dev* 2006;16:2038.
  54. Sood P, Krek A, Zavolan M, Macino G, Rajewsky N. Cell-type-specific signatures of microRNAs on target mRNA expression. *Proc Natl Acad Sci U S A* 2006;103:2746-51.
  55. Yanaihara N, Caplen N, Bowman E, Seike M, Kumamoto K, Yi M, et al. Unique microRNA molecular profiles in lung cancer diagnosis and prognosis. *Cancer Cell* 2006;9:189-98.
  56. Faraoni I, Antonetti FR, Cardone J, Bonmassar E. miR-155 gene: A typical multifunctional microRNA. *Biochim Biophys Acta - Mol Basis Dis* 2009;1792:497-505.
  57. Greither T, Grochola LF, Udelnow A, Lautenschläger C, Würl P, Taubert H. Elevated expression of microRNAs 155, 203, 210 and 222 in pancreatic tumors is associated with poorer survival. *Int J Cancer* 2010;126:73-80.
  58. Jiang S, Zhang HW, Lu MH, He XH, Li Y, Gu H, et al. MicroRNA-155 functions as an oncomiR in breast cancer by targeting the suppressor of cytokine signaling 1 gene. *Cancer Res* 2010;70:3119-27.
  59. Pedersen IM, Otero D, Kao E, Miletic AV, Hother C, Ralfkiaer E, et al. Onco-miR-155 targets SHIP1 to promote TNF $\alpha$ -dependent growth of B cell lymphomas. *EMBO Mol Med* 2009;1:288-95.
  60. Ryu JK, Hong SM, Karikari CA, Hruban RH, Goggins

- MG, Maitra A. Aberrant microRNA-155 expression is an early event in the multistep progression of pancreatic adenocarcinoma. *Pancreatology* 2010;10:66-73.
61. Roldo C, Missiaglia E, Hagan JP, Falconi M, Capelli P, Bersani S, et al. MicroRNA expression abnormalities in pancreatic endocrine and acinar tumors are associated with distinctive pathologic features and clinical behavior. *J Clin Oncol* 2006;24:4677-84.
  62. Li CL, Nie H, Wang M, Su LP, Li JF, Yu YY, et al. microRNA-155 is downregulated in gastric cancer cells and involved in cell metastasis. *Oncol Rep* 2012;27:1960-6.
  63. Rokah OH, Granot G, Ovcharenko A, Modai S, Pasmanik-Chor M, Toren A, et al. Downregulation of miR-31, miR-155, and miR-564 in chronic myeloid leukemia cells. *PLoS One* 2012;7:e35501.
  64. Dahiya N, Sherman-Baust CA, Wang TL, Davidson B, Shih LM, Zhang Y, et al. MicroRNA expression and identification of putative miRNA targets in ovarian cancer. *PLoS One*. 2008;3:e2436.
  65. Levati L, Pagani E, Romani S, Castiglia D, Piccinni E, Covaciu C, et al. MicroRNA-155 targets the SKI gene in human melanoma cell lines. *Pigment Cell Melanoma Res* 2011;24:538-50.
  66. Asangani IA, Rasheed SAK, Nikolova DA, Leupold JH, Colburn NH, Post S, et al. MicroRNA-21 (miR-21) post-transcriptionally downregulates tumor suppressor Pcdcd4 and stimulates invasion, intravasation and metastasis in colorectal cancer. *Oncogene* 2008;27:2128-36.
  67. Badalà F, Nouri-mahdavi K, Raouf DA, Meng, Fanyin, Henson, et al. MicroRNA-21 Regulates Expression of the PTEN Tumor Suppressor Gene in Human Hepatocellular Cancer. *Gastroenterology* 2007;133:647-58.
  68. Tetzlaff MT, Liu A, Xu X, Master SR, Baldwin DA, Tobias JW, et al. Differential expression of miRNAs in papillary thyroid carcinoma compared to multinodular goiter using formalin fixed paraffin embedded tissues. *Endocr Pathol* 2007;18:163-73.
  69. Tran N, McLean T, Zhang X, Zhao CJ, Thomson JM, O'Brien C, et al. MicroRNA expression profiles in head and neck cancer cell lines. *Biochem Biophys Res Commun* 2007;358:12-7.
  70. Ell B, Mercatali L, Ibrahim T, Campbell N, Schwarzenbach H, Pantel K, et al. Tumor-Induced Osteoclast miRNA Changes as Regulators and Biomarkers of Osteolytic Bone Metastasis. *Cancer Cell* 2013;24:542-56.
  71. Korpál M, Lee ES, Hu G, Kang Y. The miR-200 family inhibits epithelial-mesenchymal transition and cancer cell migration by direct targeting of E-cadherin transcriptional repressors ZEB1 and ZEB2. *J Biol Chem* 2008;283:14910-4.
  72. Erson AE, Petty EM. MicroRNAs in development and disease. *Clin Genet* 2008;74:296-306.
  73. Ceppi P, Mudduluru G, Kumarswamy R, Rapa I, Scagliotti G V., Papotti M, et al. Loss of miR-200c expression induces an aggressive, invasive, and chemoresistant phenotype in non-small cell lung cancer. *Mol Cancer Res* 2010;8:1207-16.
  74. Eulalio A, Huntzinger E, Nishihara T, Rehwinkel J, Fauser M, Izaurralde E. Deadenylation is a widespread effect of miRNA regulation. *Rna* 2009;15:21-32.
  75. Lundstrom K. Micro-RNA in Disease and Gene Therapy. *Curr Drug Discov Technol* 2011;8:76-86.
  76. Yu Z, Baserga R, Chen L, Wang C, Lisanti MP, Pestell RG. MicroRNA, cell cycle, and human breast cancer. *Am J Pathol* 2010;176:1058-64.
  77. Lima RT, Busacca S, Almeida GM, Gaudino G, Fennell DA, Vasconcelos MH. MicroRNA regulation of core apoptosis pathways in cancer. *Eur J Cancer* 2011;47:163-74.
  78. Cimmino A, Calin GA, Fabbri M, Iorio M V., Ferracin M, Shimizu M, et al. miR-15 and miR-16 induce apoptosis by targeting BCL2. *Proc Natl Acad Sci U S A* 2005;102:13944-9.
  79. Chen B, Li H, Zeng X, Yang P, Liu X, Zhao X, et al. Roles of microRNA on cancer cell metabolism. *J Transl Med* 2012;10:1.
  80. Koh HJ, Toyoda T, Fujii N, Jung MM, Rathod A, Middelbeek RJW, et al. Sucrose nonfermenting AMPK-related kinase (SNARK) mediates contraction-stimulated glucose transport in mouse skeletal muscle. *Proc Natl Acad Sci U S A* 2010;107:15541-6.
  81. Dávalos A, Goedeke L, Smibert P, Ramírez CM, Warriar NP, Andreo U, et al. miR-33a/b contribute to the regulation of fatty acid metabolism and insulin signaling. *Proc Natl Acad Sci U S A* 2011;108:9232-7.
  82. Nicoloso MS, Spizzo R, Shimizu M, Rossi S, Calin GA. MicroRNAs-The micro steering wheel of tumour metastases. *Nat Rev Cancer* 2009;9:293-302.
  83. Ma L, Weinberg RA. Micromanagers of malignancy: role of microRNAs in regulating metastasis. *Trends Genet* 2008;24:448-56.
  84. Li J, Huang H, Sun L, Yang M, Pan C, Chen W, et al. MiR-21 indicates poor prognosis in tongue squamous cell carcinomas as an apoptosis inhibitor. *Clin Cancer Res* 2009;15:3998-4008.
  85. Mayr C, Hemann MT, Bartel DP. Disrupting the Pairing Between let-7 and Hmga2 Enhances Oncogenic Transformation. *Science* 2007;315:1576-9.
  86. Koscianska E, Baev V, Skreka K, Oikonomaki K, Rusinov V, Kalantidis K. Prediction and preliminary

- validation of oncogene regulation by miRNAs. *BMC Mol Biol* 2007;8:1-14.
87. Ma L, Teruya-Feldstein J, Weinberg RA. Tumour invasion and metastasis initiated by microRNA-10b in breast cancer. *Nature* 2007;449:682-8.
  88. Huang Q, Gumireddy K, Schrier M, le Sage C, Nagel R, Nair S, et al. The microRNAs miR-373 and miR-520c promote tumour invasion and metastasis. *Nat Cell Biol* 2008;10:202-10.
  89. Sempere LF, Christensen M, Silahatoglu A, Bak M, Heath C V., Schwartz G, et al. Altered microRNA expression confined to specific epithelial cell subpopulations in breast cancer. *Cancer Res* 2007;67:11612-20.
  90. Rosenwald S, Gilad S, Benjamin S, Lebanony D, Dromi N, Faerman A, et al. Validation of a microRNA-based qRT-PCR test for accurate identification of tumor tissue origin. *Mod Pathol* 2010;23:814-23.
  91. Rosenfeld N, Aharonov R, Meiri E, Rosenwald S, Spector Y, Zepeniuk M, et al. MicroRNAs accurately identify cancer tissue origin. *Nat Biotechnol* 2008;26:462-9.
  92. Ferracin M, Veronese A, Negrini M. Micromarkers: miRNAs in cancer diagnosis and prognosis. *Expert Rev Mol Diagn* 2010;10:297-308.
  93. Gallardo E, Navarro A, Viñolas N, Marrades RM, Diaz T, Gel B, et al. miR-34a as a prognostic marker of relapse in surgically resected non-small-cell lung cancer. *Carcinogenesis* 2009;30:1903-9.
  94. Okamoto T, Miyazaki Y, Inase N. Genetic background of hypersensitivity pneumonitis. *Japanese J Chest Dis* 2010;69:701-8.
  95. Ota D, Mimori K, Yokobori T, Iwatsuki M, Kataoka A, Masuda N, et al. Identification of recurrence-related microRNAs in the bone marrow of breast cancer patients. *Int J Oncol* 2011;38:955-62.
  96. Yang H, Kong W, He L, Zhao JJ, O'Donnell JD, Wang J, et al. MicroRNA expression profiling in human ovarian cancer: miR-214 induces cell survival and cisplatin resistance by targeting PTEN. *Cancer Res* 2008;68:425-33.
  97. Weiss GJ, Bemis LT, Nakajima E, Sugita M, Birks DK, Robinson WA, et al. EGFR regulation by microRNA in lung cancer: Correlation with clinical response and survival to gefitinib and EGFR expression in cell lines. *Ann Oncol* 2008;19:1053-9.
  98. Budhu A, Jia HL, Forgues M, Liu CG, Goldstein D, Lam A, et al. identification of metastasis-related microRNAs in hepatocellular carcinoma. *Hepatology* 2008;47:897-907.
  99. Giovannetti E, Funel N, Peters GJ, Del Chiaro M, Erozcenci LA, Vasile E, et al. MicroRNA-21 in pancreatic cancer: Correlation with clinical outcome and pharmacologic aspects underlying its role in the modulation of gemcitabine activity. *Cancer Res* 2010;70:4528-38.
  100. Rodríguez-González FG, Sieuwerts AM, Smid M, Look MP, Meijer-Van Gelder ME, De Weerd V, et al. MicroRNA-30c expression level is an independent predictor of clinical benefit of endocrine therapy in advanced estrogen receptor positive breast cancer. *Breast Cancer Res Treat* 2011;127:43-51.
  101. Kota J, Chivukula RR, O'Donnell KA, Wentzel EA, Montgomery CL, Hwang H, et al. Therapeutic microRNA Delivery Suppresses Tumorigenesis in a Murine Liver Cancer Model. *Cell* 2009;137:1005-17.
  102. Iorio MV, Casalini P, Piovon C, Leva G Di, Merlo A, Triulzi T, et al. MicroRNA-205 regulates HER3 in human breast cancer. *Cancer Res* 2009;69:2195-200.
  103. Kris MG, Natale RB, Herbst RS, Lynch TJ, Prager D, Belani CP, et al. Efficacy of Gefitinib, an Inhibitor of Tyrosine Kinase, in Symptomatic Patients with Non-Small Cell Lung Cancer. *J Am Med Assoc* 2014;290:2149-58.
  104. Iorio MV, Croce CM. MicroRNA dysregulation in cancer: Diagnostics, monitoring and therapeutics. A comprehensive review. *EMBO Mol Med* 2012;4:143-59.
  105. Zhang Y, Wang Z, Gemeinhart RA. Progress in microRNA delivery. *J Control Release* 2013;172:962-74.
  106. Cho WCS. MicroRNAs: Potential biomarkers for cancer diagnosis, prognosis and targets for therapy. *Int J Biochem Cell Biol* 2010;42:1273-81.
  107. Bader AG, Brown D, Stoudemire J, Lammers P. Developing therapeutic microRNAs for cancer. *Gene Ther* 2011;18:1121-6.
  108. Si ML, Zhu S, Wu H, Lu Z, Wu F, Mo YY. miR-21-mediated tumor growth. *Oncogene* 2007;26:2799-803.
  109. Ma L, Young J, Prabhala H, Pan E, Mestdagh P, Muth D, et al. miR-9, a MYC/MYCN-activated microRNA, regulates E-cadherin and cancer metastasis. *Nat Cell Biol* 2010;12:247-56.
  110. Krützfeldt J, Rajewsky N, Braich R, Rajeev KG, Tuschl T, Manoharan M, et al. Silencing of microRNAs in vivo with "antagomirs." *Nature* 2005;438:685-9.
  111. Chen X, Liang H, Zhang J, Zen K, Zhang CY. Secreted microRNAs: A new form of intercellular communication. *Trends Cell Biol* 2012;22:125-32.
  112. Mostert B, Sieuwerts AM, Martens JWM, Sleijfer S. Diagnostic applications of cell-free and circulating tumor cell-associated miRNAs in cancer patients. *Expert Rev Mol Diagn* 2011;11:259-75.